

A (IgA) deficiency, which also precluded high-dose intravenous immunoglobulin therapy. The patient's condition deteriorated and she developed respiratory distress. A computed tomography scan showed diffuse alveolar hemorrhage (Figure 1A). Immunoabsorption (IAS) therapy using the Life 18 (Miltenyi Biotec, Bergisch Gladbach, Germany) was started with a total of 8 sessions. Treatment ameliorated thrombocytopenia and led to a resolution of the lung injury (Figure 1B). However, the patient was still dependent on dialysis. A renal biopsy revealed typical microangiopathic injury. After recurrence of pulmonary hemorrhage despite continuous high-dose methylprednisolone therapy, 10 additional daily IAS sessions were performed with clinical success. However, lung failure recurred again within 4 days after IAS withdrawal (Figure 1C) together with a rise in lactate dehydrogenase, thrombocytopenia, anemia, and a schistocyte count of 19 per mille. Thus, 4 additional sessions of IAS were necessary to control the disease again (Figure 1D). Due to low leukocyte counts and persistently low immunoglobulin levels (IgG 37 mg/dL and IgM 14 mg/dL, respectively), cytotoxic therapy was considered dangerous because of the risk for serious infections. It was, therefore, decided to administer eculizumab, a monoclonal antibody against the complement component C5, which prevents the activation of the terminal complement pathway. Within 4 days, respiratory failure

completely resolved and signs of hemolytic anemia disappeared despite cessation of IAS. Finally, therapeutic anticoagulation with low molecular heparin could be commenced. The patient was discharged dialysis dependent, but with increasing amounts of urine 71 days after admission in July 2013 with a methylprednisolone dosage of 60 mg/day and on eculizumab treatment (weekly administration of 900 mg 4 times, followed by 1200 mg fortnightly). Laboratory values at the onset of the disease, after 3 weeks, at the time of eculizumab initiation, and after achievement of stable remission are depicted in Figure 2.

Two weeks after the discharge, the patient presented again with signs of hemolysis after the fourth and fifth eculizumab infusion. In addition, the serum levels of complement C3 and C4 were consistently reduced. Serum levels of both complement components further declined in the days following eculizumab infusion (data not shown). The patient received oral anticoagulation with acenocoumarol. Eculizumab concentrations measured prior and after application revealed efficacious serum concentrations, assured complete blockage of the terminal complement pathway, and neutralizing antibodies could not be detected. Addition of mycophenolate mofetil sufficiently abrogated hemolysis, which was finally attributed to activity of the underlying SLE, indicating a lack of efficacy of eculizumab in preventing a lupus flare in this patient. The application of eculizumab could be

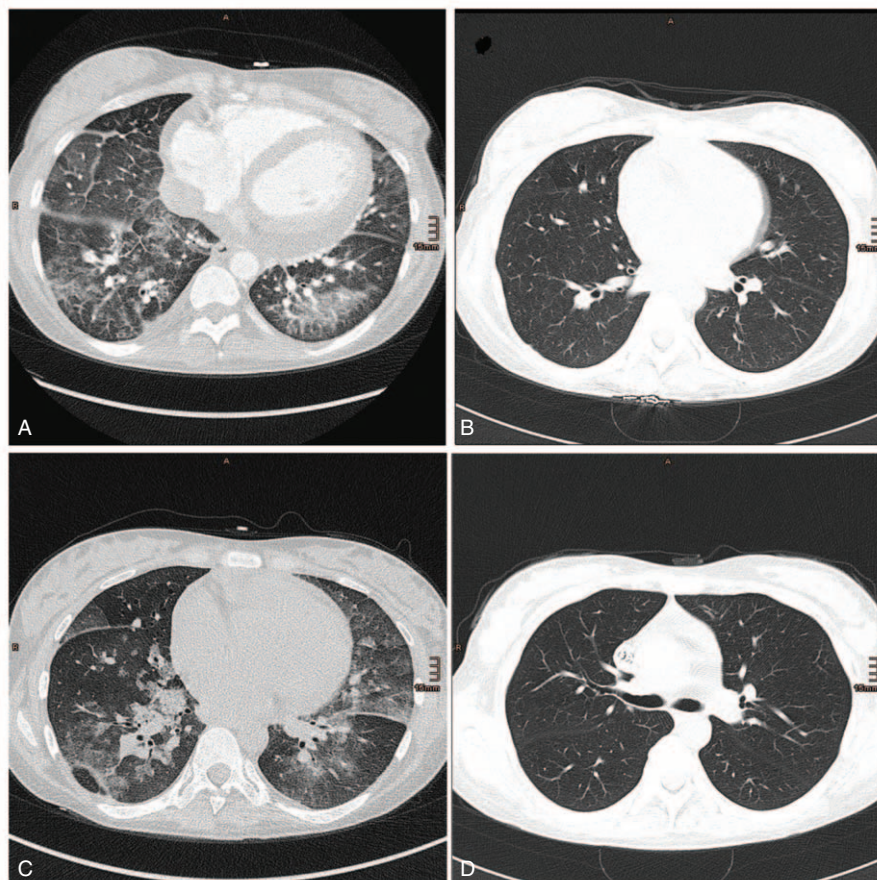


FIGURE 1. (A) Diffuse pulmonary hemorrhage in both lower lobes, which resolved after another initiation of (B) IAS. (C) After discontinuation of IAS, recurrence of pulmonary hemorrhage could be detected. These findings prompted us to initiate yet another series of IAS together with administration of eculizumab. (D) Complete resolution was detected in a control computed tomography 4 days later. IAS = immunoabsorption.

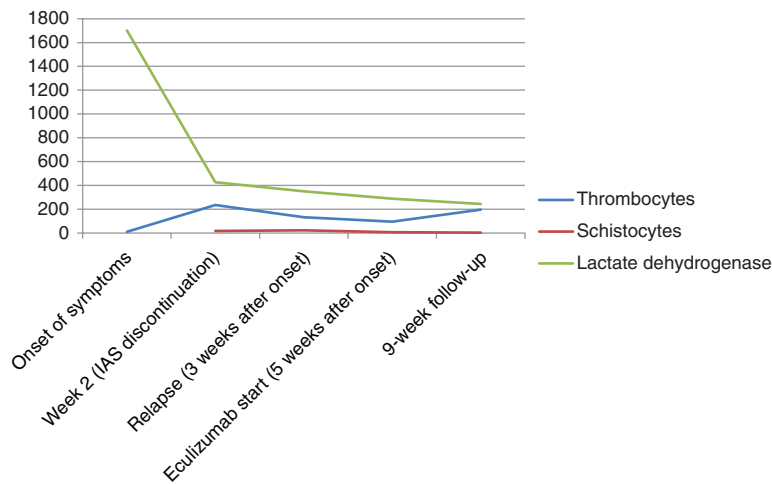


FIGURE 2. Laboratory values at the onset of the disease and at the time point of stable remission following eculizumab administration. Reference values of the respective parameters: thrombocytes (150–380 G/L), schistocytes (<5 per mille), and lactate dehydrogenase (100–250 U/L). Lactate dehydrogenase and schistocytes returned to normal values in the ninth week after onset time point and 4 weeks after initiation of eculizumab.

stopped after 3 months in September 2013 after a total of 9 infusions without recurrence of thrombotic microangiopathy despite a persisting positive test for antiphospholipid antibodies. Several measurements of C3d and the terminal complement complex (sC5bC9) while under treatment revealed normal serum levels (data not shown). Examination of the complement regulatory protein factors H and I as well as the alternative pathway component factor B revealed values within a normal range, indicative of a normal alternative complement pathway. However, persistently reduced C3 and C4 serum levels along with a low C1q (normal in one measurement and slightly reduced in another) are suspicious for a consumption of the classical complement pathway (Table 1). In addition, to exclude inherited predisposing factors in complement C3 or C4, genetic analysis was performed. A rare heterozygous synonymous variation in exon 13 of the *C3* gene (c.1677C>T; p.C559C) was detected, which was predicted as disease causing by MutationTaster ([\[www.mutationtaster.org/\]\(http://www.mutationtaster.org/\)\) with a high probability \(0.99\) \(Table 2\). Analysis of *C4* revealed the presence of 2 copies of the *C4A* and 1 copy of the *C4B* genes \(data not shown\).](http://</p></div><div data-bbox=)

One year after the initial insult, the patient is still dialysis dependent. Steroid reduction was well tolerated without signs of active thrombotic microangiopathy. The patient is listed for renal transplantation. Her current medication consists of mycophenolate mofetil 250 mg twice a day, methylprednisolone 10 mg every second day, blood pressure-lowering medication, diuretics, thyroid hormone substitution, and acenocoumarol with a target international normalized ratio of 2 to 3.

DISCUSSION

Eculizumab has already shown encouraging results in a patient with recurrence of CAPS with reversal of thrombocytopenia and prevention of further clinical episodes of

TABLE 1. Complement Analysis After Application of Eculizumab and 6 mo After Cessation of Therapy

	On Treatment With Eculizumab (09/2013)	Off Treatment With Eculizumab (02/2014)
Classical pathway (total complement activity) (range 48–103 CH50/mL)	0	39
Alternative pathway (total complement activity) (70%–105%)	1	67
Mannose-binding lectin pathway (0%–110%)	1	Not examined
C1q antigen (66–180 mg/L)	76	65
C2 (600–4000 CH63/mL)	1790	Not examined
C3 (0.9–1.8 g/L)	0.77	0.57
C3a (range 70–270 U/mL)	168	149
C4 (0.15–0.55 g/L)	0.07	0.09
sC5b9 (terminal pathway activation, range 110–252 U/L)	179	311
Factor H antigen (127–447 mg/L)	367	545
Factor I antigen (70%–130%)	88	85
Factor B antigen (70%–130%)	93	87
Anti-C1q autoantibody	Negative	
Anti-factor H IgG autoantibody	Negative	

IgG = immunoglobulin G.

TABLE 2. Genetic Analysis of Complement C3

C3						
Exon	Protein	cDNA position	SNP ID according to dbSNP	Type	Genotype	Minor allele
13	p.C559C	c.1677C>T	—	Coding-synonymous	Heterozygous	T
Tool	Qualitative prediction		Quantitative prediction/ Probability of prediction		Explanation	
MutationTaster	Disease causing		Probability: 0.99		The MutationTaster probability value is the probability of the prediction, that is, a value close to 1 indicates a high “security” of the prediction.	

cDNA and protein positions are indicated according to transcript (NM_000064.2) and protein (NP_000055.2), respectively. The prediction of the functional relevance of p.C559C was performed by MutationTaster (<http://www.mutationmaster.org/>).

thrombosis.⁵ Its successful use has also been reported in recurrence of CAPS after renal transplantation.^{6,7} Transplantation in CAPS is also possible with prophylactic eculizumab administration.⁸ In addition, murine models reveal a pivotal role of the complement system in antiphospholipid antibodies-induced thrombosis along with endothelial cell and platelet activation. Prevention of terminal complement formation by using a monoclonal antibody against C5 inhibited thrombophilia induced by antiphospholipid antibodies.⁹ Complement activation and consumption was also confirmed in our patient by the finding that serum levels of C3 and C4 were significantly lowered while disease was highly active. Immunohistochemical analysis of the kidney biopsy specimen revealed a strong staining for C1q and IgM, while deposition of C3, IgG, and IgA was sparsely present. Interestingly, administration of eculizumab was associated with a further decrease in C3 and C4 serum levels. Moreover, sufficient eculizumab levels and complement inhibition was not capable of preventing a flare of her underlying SLE. Evidence that eculizumab may not be effective in SLE is still lacking, since there is limited data coming from a phase I single-center trial. No changes in laboratory values and systemic lupus erythematosus disease activity index have been observed in this preliminary clinical trial. However, the cohort of patients had low disease activity, thus precluding considerations on the therapeutic efficacy of eculizumab.¹⁰

Our case confirms previous reports that it is possible to discontinue eculizumab in CAPS after complete remission despite a continuing positive test for antiphospholipid antibodies. Persistently, reduced C4 and C3 levels in our patient can be explained by continuous activation of the classical complement pathway induced by immune complexes and antiphospholipid antibodies that cannot be influenced by eculizumab. Furthermore, a lowered C1q serum level after eculizumab cessation and strong deposition in the kidney biopsy also supports a consumption of the classical complement pathway. The finding that eculizumab application was followed by a further decrease in C4 and C3 serum levels is intriguing. One possible explanation could be that eculizumab interfered with clearance of immune complexes and thus caused a further stimulation of the classical pathway. Genetic analysis revealed a rare heterozygous variation in exon 13 of the C3 gene. This mutation does not cause an amino acid change in the C3 protein, so it is likely not causative; however,

a potential influence cannot be excluded, as its predicted functional relevance showed a high probability as a disease-causing factor.

CAPS remains a severe variant of the APS with a high mortality rate of approximately 50%.¹ However, patients achieving disease control are prone to a persisting morbidity as reported herein. This case is unique since the patient presented with a continuum of autoimmune disorders and had underlying IgA deficiency. Due to the latter, the standard therapy for CAPS was contraindicated or not tolerated by the patient. The patient had a good clinical response toward IAS combined with high doses of methylprednisolone, whereas she was refractory to rituximab. Inhibition of the terminal complement pathway by eculizumab led to a persistent clinical remission without evident recurrence of disease.

CONCLUSION

Targeting complement activation may provide a new therapeutic option in the treatment of CAPS, especially in patients refractory or unable to tolerate standard therapy. In our exceptional case, stabilization of the disease course was possible after eculizumab was commenced. Further clinical investigations are necessary to clarify whether it can be generalized that blockade of the terminal complement activation is efficacious in the treatment of CAPS.

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