

# Loss of Estrogen Receptor Beta Expression Correlates with Shorter Overall Survival and Lack of Clinical Response to Chemotherapy in Ovarian Cancer Patients

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**Abstract.** *Background:* Estrogen receptor beta (ER $\beta$ ) belongs to a large family of nuclear receptors. Recent studies have suggested that ER $\beta$  in contrast to ER $\alpha$  might act as a tumour suppressor in ovarian cancer (OVCA). *Materials and Methods:* Expression of ER $\beta$  was detected by immunocytochemistry in 11 OVCA cell lines and by immunohistochemistry in 43 (41 FIGO stage III) OVCA specimens prepared before chemotherapy and 30 specimens from the same group after chemotherapy. Cisplatin sensitivity in the 11 cell lines was also analysed. *Results:* No significant correlations between cisplatin-sensitivity and expression of ER $\beta$  was found in the cell lines. In the cases which responded well to chemotherapy (complete response) ER $\beta$  expression at preliminary laparotomy (PL) was significantly higher ( $p=0.0004$ ) than in those with progressive disease. Kaplan-Meier analysis revealed that the patients with higher ER $\beta$  expression (>30% of cells) at PL had an increased overall survival time and progression-free time ( $p=0.00161$  and  $p=0.03255$ , respectively) than the patients with lower ER $\beta$  expression. Significantly shorter overall survival time characterized the cases with lower immunoreactivity score of ER $\beta$  expression at secondary

cytoreduction (SCR) ( $p=0.00346$ ). *Conclusion:* The loss of ER $\beta$  expression in ovarian tumours may be a feature of malignant transformation.

Ovarian cancer (OVCA) is one of the most lethal gynaecological carcinomas worldwide. About 190,000 new cases and 114,000 deaths from ovarian cancer are estimated to occur annually. The highest rates are reported in Scandinavia and Eastern Europe, the USA and Canada (1). Because early-stage OVCA (FIGO I or II) is generally asymptomatic, approximately 75% of women present with advanced disease at diagnosis which is associated with poor prognosis. Survival is highly dependent on the stage of the disease: 5-year survival in patients with early-stage is 80-90% compared to 25% for patients with advanced OVCA (2).

High serum levels of estrogen have been implicated as a risk factor for OVCA, but the cellular signal transduction pathways involved are not completely well known. The incessant ovulation hypothesis argues that trauma and repair of ovarian epithelium induced ovulation, contributes to OVCA development. Ovulatory cycles lead to long-term exposure of the epithelium to an estrogen-rich environment, which may promote cellular proliferation, including cyst formation and possibly malignant transformation (3).

Estrogen receptors, ER alpha (ER $\alpha$ ) and ER beta (ER $\beta$ ) belong to a large family of nuclear receptors and mediate the action of estrogens as ligand-dependent transcription factors (4). The molecular mechanisms of ER $\beta$  function in OVCA are still poorly established, but recent studies have suggested that ER $\beta$  in contrast to ER $\alpha$  might act as a tumour

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*Key Words:* Estrogen receptor beta, ovarian cancer, prognostic factor, immunohistochemistry, cisplatin.

suppressor in OVCA (4, 5). Preliminary studies have revealed that ER $\beta$  levels, both protein and mRNA in OVCA decline relative to levels in the normal ovary (4, 6-8). Chan *et al.* (9), demonstrated that the immunoreactivity score of ER $\beta$  expression was significantly higher in normal tissue compared with malignant and its expression was a significant good predictor for disease-free survival and overall survival. Furthermore, the absence of ER $\beta$  in OVCA might be a feature of malignant transformation (9).

From the 1970s, the ERs has evolved to be the most effective target for breast and ovarian cancer therapy. Interactions between estradiol (E2) and ERs can be effectively blocked using a variety of agents, such as selective ER modulators (SERMs). Tamoxifen and raloxifen as a leading SERMs, are competitive inhibitors of E2 at the ERs and display distinct effects depending on the tissues (10). Only 15-18% of ER-positive OVCA initially respond to anti estrogen treatment based on blocking of estrogen-ER binding, in contrast to effective treatment of about 50% of ER-positive breast carcinomas (11). The most common mechanism of antiestrogen resistance is the absence of ER. From the previous data, the role of tamoxifen in OVCA has not been properly established, although some authors have suggested that combined therapy with cisplatin and tamoxifen might reduced cisplatin resistance (12, 13).

The significance of ER $\beta$  expression in ovarian carcinogenesis and its impact on growth and survival of OVCA cells is still controversial. In this study, the expression of ER $\beta$ , in malignant epithelial ovarian tumours, and OVCA cell lines was investigated. Because cisplatin resistance is a major obstacle in the treatment of OVCA, analysis of cisplatin sensitivity in 11 OVCA cell lines was also performed.

## Materials and Methods

**Cell culture.** The cisplatin-resistant cell line, A2780RCIS, was derived from the OVCA cell line, A2780 (14). The human OVCA cell lines CAOV-3, EFO 21, EFO 27, ES-2, MDAH 2774, OAW 42, OVCAR-3, PA-1 and SKOV-3 were kindly provided by Dr. Carsten Denkert (Institute of Pathology, Charité, Berlin, Germany). The human OVCA cells were grown in Leibovitz L-15 medium (Biowhittaker, Walkersville, MD, USA) supplemented with 10% fetal calf serum (FCS) (GIBCO/BRL, Grand Island, NY, USA), 1 mM L-glutamine, 6.25 mg/l fetuin, 80 IE/l insulin, 2.5 mg/ml transferrin, 0.5 g/l glucose, 1.1 g/l NaHCO<sub>3</sub>, 1% minimal essential vitamins and 20,000 kIE/l trasylol in a humidified atmosphere of 5% CO<sub>2</sub> at 37°C as described previously (14-17). In order to ensure maintenance of the cisplatin-resistant phenotype of the A2780RCIS cells, the medium was supplemented with 10 Ag/mL of cisplatin (33.3  $\mu$ mol/L; GRY-Pharm, Kirchzarten, Germany).

**Cell proliferation assay.** Chemoresistance was tested using a proliferation assay based on sulphorhodamine B (SRB) staining as described previously (18). Briefly, 800 cells per well were seeded in 96-well plates in triplicate. After 24 h attachment, cisplatin was

added in dilution series for a 5-day incubation, before SRB staining was performed. Incubation with cisplatin was terminated by replacing the medium with 10% trichloroacetic acid, followed by incubation at 4°C for 1 h. Subsequently, the plates were washed five times with water and stained by adding 100  $\mu$ l 0.4% SRB (Sigma, St. Louis, MO, USA) in 1% acetic acid for 10 min at room temperature. Washing the plates five times with 1% acetic acid eliminated unbound dye. After air-drying and re-solubilization of the protein bound dye in 10 mM Tris-HCl (pH=8.0) absorbance was read at 562 nm in an Elisa-Reader (EL 340 Microplate Bio Kinetics Reader, BIO-TEK Instruments, Winooski, VT, USA). The measurements were performed in triplicates in three independent experiments. The IC<sub>50</sub>-values were calculated from three independent experiments for each cell line.

**Patients.** Immunohistochemical examination was performed retrospectively on tissue samples taken for routine diagnostic purposes. Forty three patients who had undergone surgery in 1999-2002 due to OVCA in the Department of Gynaecology and Obstetrics, University Medical School in Poznań, Poland were included in the study. The cases were selected based on availability of tissue and were not stratified for known preoperative or pathological prognostic factors. The study was approved by an Institutional Review Board (IRB) and the patients gave their informed consent before their inclusion into the study. Following the primary laparotomy (PL) all the patients were subjected to chemotherapy using cisplatin-based schemes (Table I). Thirty six patients from the same group were also subjected to secondary cytoreduction (SCR). In seven cases no second-look procedure was performed due to advancement of the disease. In six cases no tumour cells were detected in the material originating from the second-look procedure. The patients were monitored by periodic medical check-ups, ultrasonographic, radiological and cancer antigen 125 (CA-125) serum levels examinations. During the 52 months follow-up period, 22 patients (51%) had recurrent disease and 13 patients (30%) died of the disease. The mean progression-free survival time was 16.9 months (range 0 to 52 months), while the mean overall survival time was 24.6 months (range 6 to 52 months). Only the one stage I and one stage II patient achieved optimal cytoreduction.

Tissue samples were fixed in 10% buffered formalin and embedded in paraffin. In each case, hematoxylin and eosin stained preparations were subjected to histopathological evaluation by two pathologists. The stage of the tumours was assessed according to the International Federation of Gynaecology and Obstetrics (19). Tumours were graded according to the Silverberg grading system (20).

**Immunohistochemistry.** Formalin-fixed, paraffin embedded tissue was freshly cut (4  $\mu$ m). The sections were mounted on Superfrost slides (Menzel Gläser, Braunschweig, Germany), dewaxed with xylene and gradually hydrated. Activity of endogenous peroxidase was blocked by 5 min exposure to 3% H<sub>2</sub>O<sub>2</sub>. All the sections were boiled for 15 min at 250W in Antigen Retrieval Solution (DakoCytomation, Glostrup, Denmark). Then, immunohistochemical reactions were performed using the mouse mAb PPG5/10 (DakoCytomation) directed against ER $\beta$  1 (dilution 1:50 in Antibody Diluent, Background Reducing (DakoCytomation) 1 h at 20°C). Each reaction was accompanied by a negative control using Primary Mouse Negative Control (DakoCytomation). Subsequent incubations involved biotinylated antibodies (15 min, room temperature) and streptavidin-biotinylated peroxidase complex (15

Table I. Patient and tumour characteristics.

Patient and tumour characteristics	N <sup>c</sup>	%
All patients	43	100
Age (mean 51) <sup>a</sup>		
≤50	20	47
50-60	16	37
>60	7	16
Grade <sup>a</sup>		
1	7	16
2	18	42
3	18	42
FIGO <sup>a</sup>		
I	1	2
II	1	2
III	41	95
Histology <sup>a</sup>		
Serous	37	86
Endometrioid	3	7
Other	3	7
Clinical response <sup>b</sup>		
Complete response	16	37
Stable disease	5	12
Progressive disease	22	51
Chemotherapy (in total)		
Cisplatin/Paclitaxel	31	72
Cisplatin/Cyclophosphamide/Adriablastin	6	14
Cisplatin/Cyclophosphamide/Paclitaxel	3	7
Cisplatin/Cyclophosphamide/Paclitaxel/Adriablastin	2	5
Carboplatin/Paclitaxel	1	2

<sup>a</sup>Data are given for the first operation/diagnosis implemented.

<sup>b</sup>According to RECIST 1.0 (Response Evaluation Criteria in Solid Tumours) (29). <sup>c</sup>Differences in the sum to 100 % in groups are due to rounding.

min, room temperature) (Dako Detection System, LSAB+ and horseradish peroxidase, HRP, DakoCytomation). Diaminobenzidine (DAB, DakoCytomation) was used as a chromogen (7 min, at room temperature). All the sections were counterstained with Meyer's hematoxylin. In this study, the Ki67 (21), and p53 (22) expression data, which had been investigated previously on the same group of patients (23), were reused.

**Immunocytochemistry.** Immunostaining of ERβ was performed in all the cell lines. The cells were grown on microscope slides and fixed in on ice-cold methanol-acetone mixture (1:1) for 10 min. After re-hydration, the immunostaining reaction was performed in triplicate as described above. Furthermore, immunostaining of Ki67 was conducted for all the cell lines with mouse mAB against Ki67 (clone MIB-1; dilution 1:100 in Antibody Diluent, Background Reducing, DakoCytomation, 1 h at 20°C).

**Evaluation of reaction intensity.** The intensity of the immunohistochemical and immunocytochemical reactions were estimated independently by two pathologists. Assessment of staining was "blinded" to sample details. In doubtful cases a re-evaluation was performed using a double-headed microscope and staining was discussed until a consensus was achieved. Immunostaining reactions

Table II. Chemosensitivity to cisplatin (IC<sub>50</sub> value) and immunoreactivity score of ERβ and Ki67 expression in human ovarian carcinoma cell lines.

Cell line	ERβ expression nuclear	ERβ expression cytoplasmic	Ki67 expression <sup>a</sup>	Cisplatin IC <sub>50</sub> (μM)
A2780P	0	0	20	23.87
A2780RCIS	0	0	20	98.98
CAOV-3	0	0	60	1.92
EFO 21	3	0	10	5.08
EFO 27	0	0	15	2.25
ES-2	0	1	80	7.64
Mdah 2774	0	0	90	6.36
OAW 42	0	0	5	5.49
OVCAR-3	0	2	20	1.88
PA-1	0	0	80	0.75
SKOV-3	0	0	75	18.85

<sup>a</sup>Percentage of positive cells.

of p53 and Ki67 were evaluated using percentage of positive cells score. The expression of ERβ was graded using a semiquantitative scoring scale based on percentage of reactive cells (no staining=score 0, <10%=score 1, 10-30%=score 2, >30%=score 3). Consequently, four possible products (0, 1, 2, 3) were obtained.

**Statistical analysis.** Statistical analysis of the results took advantage of Statistica 98 PL software (Statsoft, Krakow, Poland), SPSS software (release 10.0; SPSS Inc., Chicago, IL, USA) and Microsoft Office Excel 2007 (Microsoft Corporation, Redmond, WA, USA). The employed tests included Mann-Whitney *U*-test, Spearman's rank correlation, Chi-square and Kruskal-Wallis test. Kaplan-Meier statistics and log-rank tests were performed to estimate the significance of differences in survival times. The length of progression-free survival was defined as the time between the primary surgical treatment and diagnosis of a recurrent tumour or death.

## Results

**Cisplatin-sensitivity of cell lines.** The sensitivity of the various human OVCA cell lines against treatment with cisplatin was determined by the assessment of IC<sub>50</sub> values (Table II). The most sensitive cell line, PA-1, exhibited an IC<sub>50</sub> of 0.75 μM, and the most cisplatin-resistant cell line, A2780RCIS, demonstrated an IC<sub>50</sub> of 98.98 μM.

**ERβ immunostaining in cell lines.** Strong nuclear ERβ expression (score 3) was shown in the EFO 21 cells (Figure 1A). Interestingly, the immunocytochemical experiments showed that ERβ could also be detected in the cytoplasm of the OVCAR-3 and ES-2 cells (score 2 and 1, respectively). The other cell lines were ERβ negative (Figure 1B, EFO 27 cells). No significant correlations were found between cisplatin-sensitivity and the expression of ERβ in the OVCA

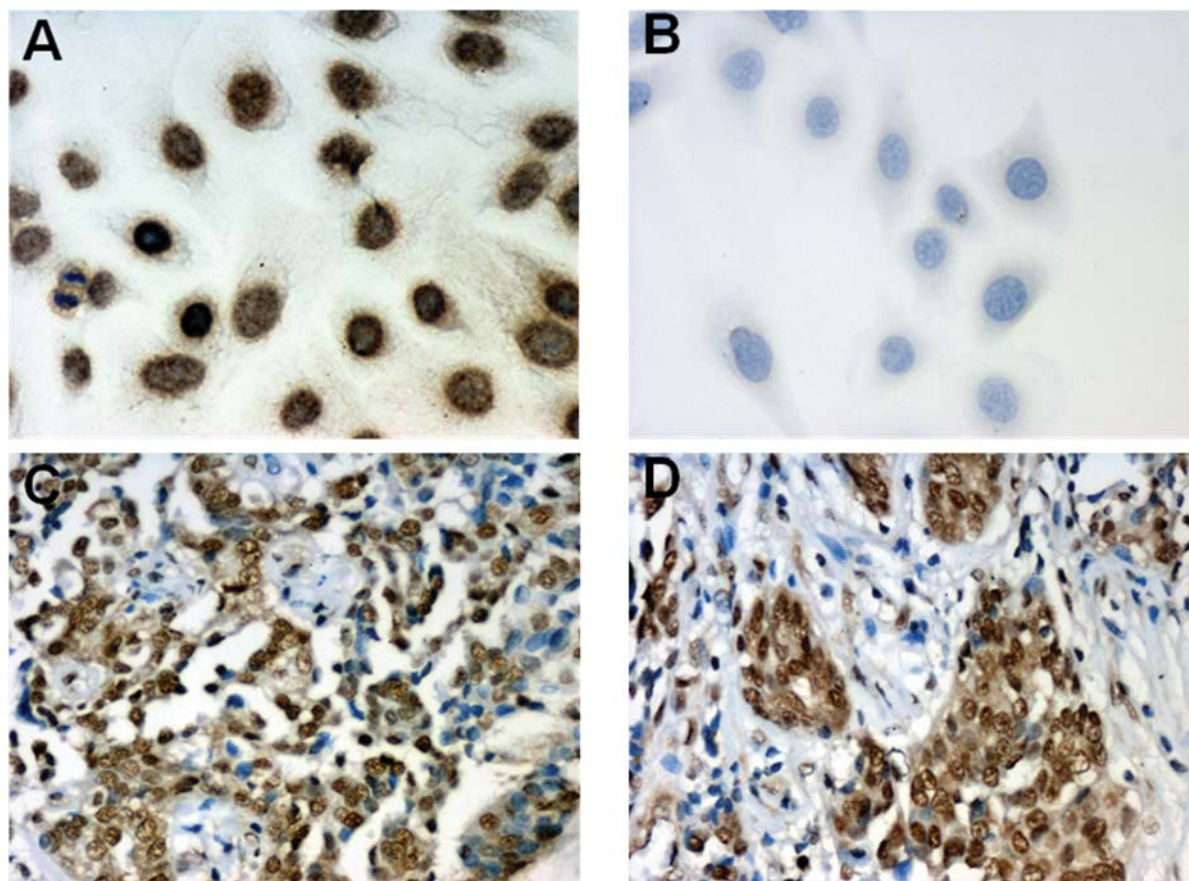


Figure 1. Immunohistochemical localization of ERβ expression in EFO 21 cells (A., ×200; hematoxylin), EFO 27 cells (B., ×200; hematoxylin) and in ovarian cancer tissue (C., ×200, D., ×200; hematoxylin).

cell lines. Additionally, statistical analysis did not reveal any correlations between the expression of ERβ and Ki67. Figures 2A and 2B present the expression of Ki67 in Mdah 2774 and OVCAR-3 cells, respectively.

*ERβ expression and clinicopathological and immunohistochemical parameters.* Immunoreactivity of variable intensity was obtained in the individual OVCA cases (Figure 1C and D). At the first stage of statistical analysis of relationships between ERβ expression and clinicopathological parameters of the patients, the Kruskal-Wallis test was used. The relationship between the percentage of positive ERβ cells and the histological type of the tumour, the grade and the clinical response to chemotherapy was examined (Figure 3, Table III). In the cases which responded well to chemotherapy (complete response) based on cisplatin the immunoreactivity score of ERβ expression at PL was significantly higher ( $p=0.0004$ ) than in patients with progressive disease (Figure 4). Statistical analysis did not reveal any significant correlations between ERβ expression and Ki67 and p53 percentage of positive cells (Table III).

Table III. Correlation between estrogen receptor beta expression and various clinicopathologic and immunohistochemical parameters.

Characteristics	p-Value
Histological type <sup>a</sup>	0.6881
Grade <sup>a</sup>	0.5379
Clinical response <sup>a</sup>	0.0004
Age <sup>b</sup>	0.7932
Ki67 <sup>b</sup>	0.2742
p53 <sup>b</sup>	0.1647
CA-125 <sup>b</sup>	0.7485

<sup>a</sup>ANOVA rang Kruskal-Wallis. <sup>b</sup>Spearman's rank correlation.

In the Kaplan-Meier analysis overall survival time and progression-free time were compared between cases showing lower (0-30%) or higher (31-100% of positive cells) immunoreactivity score of ERβ expression at PL and SCR. The patients with higher ERβ expression at PL had an increased overall survival time (Figure 5A) and an increased

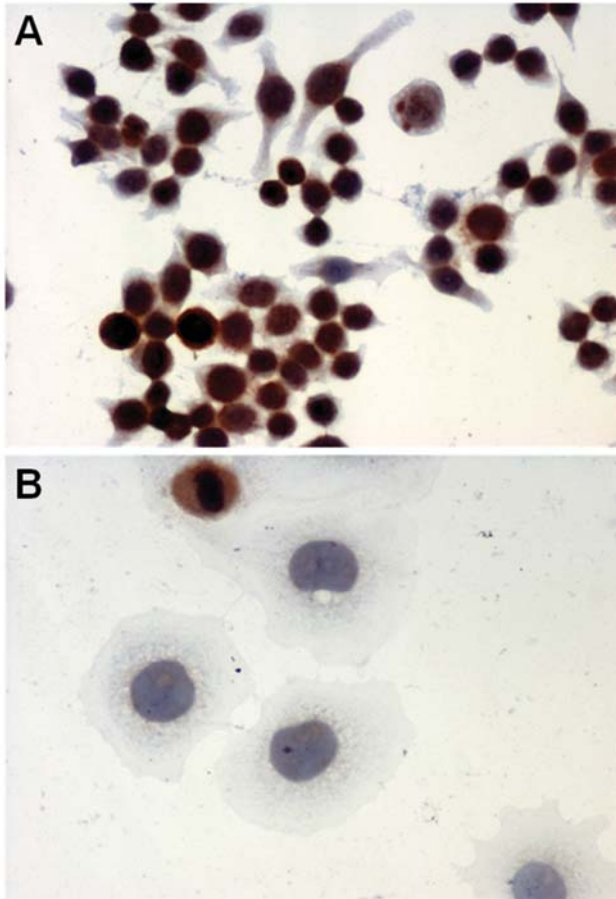


Figure 2. Immunocytochemical localization of Ki67 expression in Mdah 2774 cells (A., ×200; hematoxylin) and OVCAR-3 cells (B., ×400; hematoxylin).

progression-free time (Figure 5B) than the patients with lower ERβ expression. The analysis also demonstrated that significantly shorter overall survival time characterized the cases with lower immunoreactivity score of ERβ expression at SCR (Figure 6A). No significant differences in progression-free time between the patients with lower and higher ERβ expression at SCR was observed (Figure 6B).

Since no significant relationships between the studied clinicopathological parameters (age, histology, grade, CA-125 at PL level) and overall survival and progression-free time was found with univariate analysis ( $p>0.05$ ), a multivariate analysis was not performed. Since 95% of the patients were in stage FIGO III, relationships between stage and survival data were not investigated.

**Discussion**

The most sensitive OVCA cell line, PA-1, exhibited an IC<sub>50</sub> of 0.75 μM and the most cisplatin-resistant cell line, A2780RCIS, demonstrated an IC<sub>50</sub> of 98.98 μM. However

ERβ (%)		Ki 67 (%)	p53 (%)	Grade	Response	Recurrence	Death
0-30%	0	60	20	1	PD	1	0
	0	70	60	1	PD	1	1
	0	60	90	2	PD	1	1
	0	80	20	2	PD	1	1
	0	70	90	2	PD	1	1
	0	80	70	2	PD	1	1
	0	25	60	3	PD	1	0
	0	60	70	3	PD	1	0
	0	40	90	3	PD	1	1
	5	30	15	1	CR	0	0
	5	60	0	1	PD	1	1
	5	30	40	3	PD	1	1
	5	60	90	3	PD	1	1
	5	90	90	3	PD	1	1
	10	90	60	2	CR	0	0
	10	30	0	2	PD	1	1
	10	80	80	3	PD	1	0
	10	80	90	3	PD	1	1
	15	50	25	3	SD	0	0
	20	60	5	2	CR	0	0
	20	80	70	3	CR	0	0
	20	90	90	3	PD	1	1
	25	30	0	1	SD	0	0
	25	80	40	2	CR	0	0
	25	40	15	2	SD	0	0
31-100%	40	50	15	2	SD	0	0
	40	80	30	2	PD	1	0
	40	60	90	2	PD	1	0
	40	30	5	3	CR	0	0
	50	20	20	1	CR	0	0
	50	70	15	2	SD	0	0
	50	30	30	3	CR	0	0
	60	30	5	1	CR	0	0
	60	60	15	2	PD	1	0
	60	90	30	3	PD	1	0
	70	20	90	2	CR	0	0
	80	70	70	2	CR	0	0
	80	30	5	2	CR	0	0
	80	40	80	3	CR	0	0
	80	70	0	3	CR	0	0
80	50	90	3	CR	0	0	
80	90	90	3	PD	1	0	
90	10	15	2	CR	0	0	

Figure 3. Expression of ERβ at preliminary laparotomy in relation to clinical and pathological data of the patients. CR: complete response, SD: stable disease, PD: progressive disease.

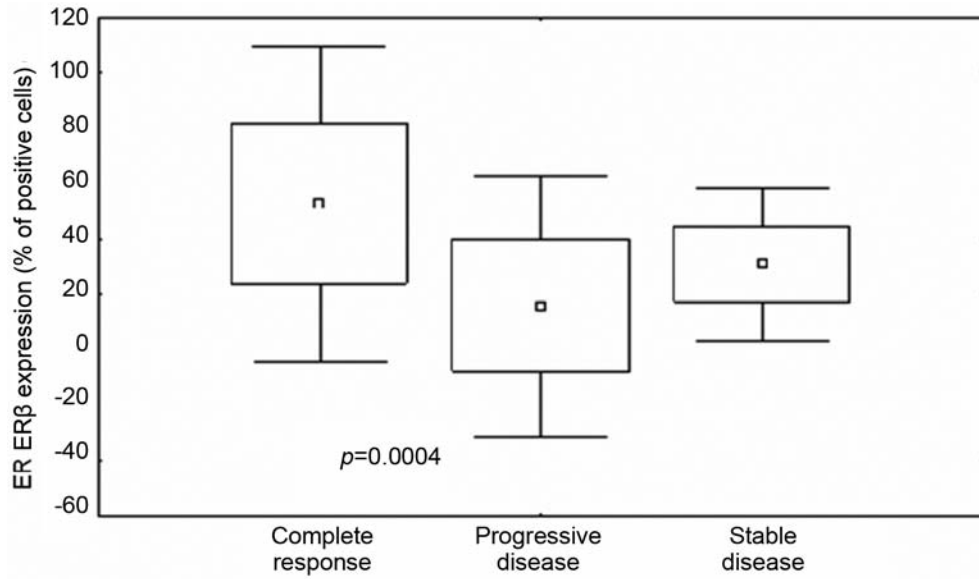


Figure 4. Expression of ERβ and clinical response to chemotherapy.

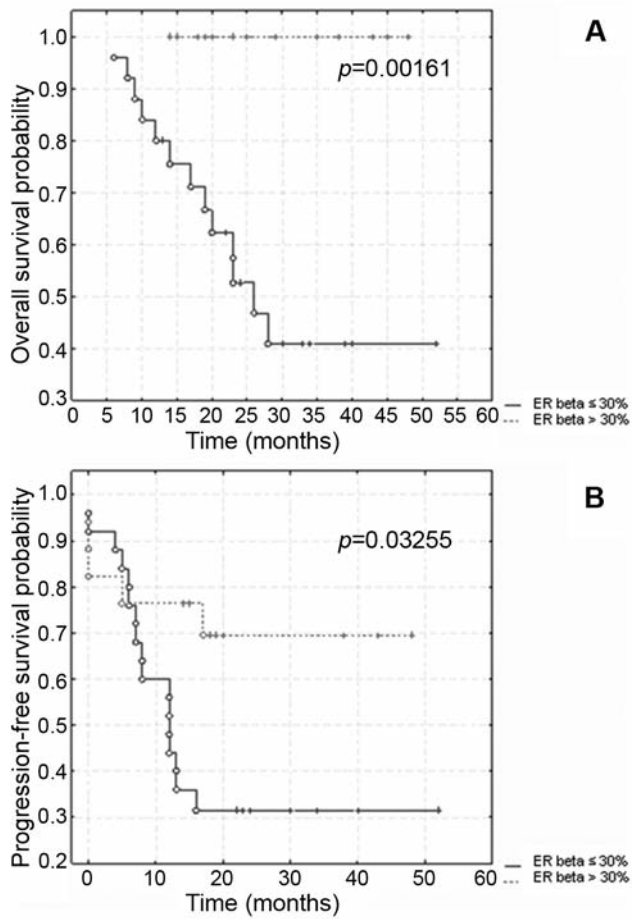


Figure 5. Kaplan-Meier curves for survival in relation to ERβ expression at preliminary laparotomy.

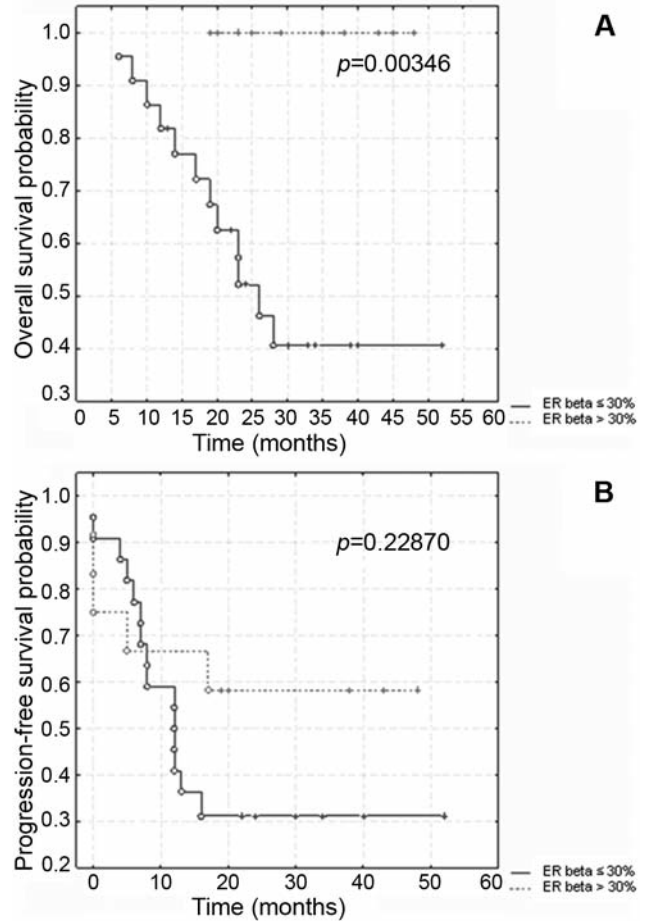


Figure 6. Kaplan-Meier curves for survival in relation to ERβ expression at secondary cytoreduction.

no significant correlations between cisplatin-sensitivity and ER $\beta$  expression were found in the cell lines, nor were any correlations between expression of ER $\beta$  and Ki67 found. Treeck *et al.* (24) studied the role of ER $\beta$ 1 and two of its splice variants (ER $\beta$ - $\delta$ 125, ER $\beta$ - $\delta$ 1256) in the regulation of gene expression, apoptosis, cellular proliferation and migration of SKOV-3 OVCA cells and demonstrated that ER $\beta$ 1 expression displayed multiple antitumoral effects.

The expression of ER $\beta$  in the tissue samples from OVCA patients was not correlated with the histopathological parameters, such as histological type and cancer grade. In the cases which responded well to chemotherapy based on cisplatin the immunoreactivity score of ER $\beta$  expression at PL was significantly higher ( $p=0.0004$ ) than in the patients with progressive disease. Moreover, Kaplan-Meier analysis revealed that the patients with higher ER $\beta$  expression at PL had an increased overall survival time ( $p=0.00161$ ) and an increased progression-free time ( $p=0.03255$ ) than the patients with lower ER $\beta$  expression. Significantly shorter overall survival time also characterized the cases with lower of ER $\beta$  expression at SCR ( $p=0.00346$ ).

The loss of ER $\beta$  expression in ovarian tumours may be a feature of malignant transformation. Chan *et al.* (9) in addition to showing that ER $\beta$  expression was significantly higher in normal ovary than malignant tissues, also showed significant correlations between the stage of disease and level of ER $\beta$  expression (higher in stage I than stage II-IV). Multivariable analysis confirmed the role of ER $\beta$  in ovarian carcinogenesis, because its higher expression was found to be connected with longer disease-free survival as well as overall survival as confirmed in present study.

Several earlier studies have also investigated the status of ER status in relation to survival (25, 26). Interestingly, Fujimoto *et al.* (26) found that patients with a low or high ratio of ER $\alpha$  to ER $\beta$  had a significantly worse prognosis than patients with a more equal ratio.

In metastatic OVCA cells ER $\beta$  expression is often lost (27) however, its role in ovarian carcinogenesis is still unclear. EMT (epithelial-mesenchymal transition), a highly conserved process in which epithelial cells undergo a phenotypic switch to form mesenchymal-like cells, involves the loss of polarity and intracellular adhesion. These molecular and structural alterations are thought to allow dynamic cellular migration and the development of local and distant metastases. Park *et al.* identified ER $\beta$  overexpression as an important factor in the inhibition of prometastatic effects connected with the promotion of EMT in OVCA cells (28). Their results suggested that ER $\beta$  may have a function as a negative modulator of ER $\alpha$  in carcinogenesis of the ovary and the development of metastatic potential.

Overall, greater understanding of the role of ER $\beta$  expression in ovarian cancer may help to develop and

explore further the potential of hormonal therapy in ovarian tumours. Future studies on ER $\beta$  are needed to determine fully the clinicopathological implications of ER $\beta$  in ovarian carcinogenesis.

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*Received October 31, 2010*

*Revised January 23, 2011*

*Accepted January 25, 2011*