

RESEARCH

Open Access

Circulating survivin levels in healthy and asthmatic pregnancy

Andras Bikov^{1*}, Renata Bocskei¹, Noemi Eszes¹, Aniko Bohacs¹, Gyorgy Losonczy¹, Janos Rigo², Ildiko Horvath¹ and Lilla Tamasi¹

Abstract

Background: Asthma is one of the most common conditions which complicate pregnancy. Pro- and anti-apoptotic mechanisms can be modulated by asthma accompanying pregnancy. Survivin, an anti-apoptotic protein has been implicated in the pathomechanism of asthma and also in the development of pathological pregnancies; however survivin has not been studied in pregnant asthmatics.

Methods: Twenty-eight asthmatic pregnant (AP), 25 asthmatic non-pregnant (ANP), 21 healthy pregnant (HP) and 29 healthy non-pregnant (HNP) women were enrolled in this cross-sectional study. Plasma survivin concentration was determined by ELISA.

Results: Plasma survivin was significantly lower in HP (1.64 /0-74.9/ pg/ml) than in HNP (24.6 /0-333.3/ pg/ml, $p = 0.01$). However, this difference was not observed between the asthmatic groups ($p = 0.64$). Similarly, there was no difference either between HNP and ANP (10.5 /0-215.4/ pg/ml, $p = 0.23$) or between HP and AP (13.9 /0-364.1/ pg/ml, $p = 0.30$) groups.

Conclusions: Decreased plasma survivin levels in physiological but not in asthmatic pregnancy may suggest that the normal apoptotic mechanisms are compromised in asthmatic gestation.

Keywords: Apoptosis, Asthma, Pregnancy, Survivin

Background

Asthma is one of the most common disorders which may complicate pregnancy and it represents an increased risk for maternal and foetal complications, including pre-eclampsia, gestational hypertension, preterm delivery, Caesarean section, low birth weight, intrauterine growth restriction and foetal death [1,2]. Asthma complicates 4-8% of pregnancies [2]. In addition, it is estimated that one third of asthmatic women experience asthma worsening during gestation [3]. The natural course of asthma during pregnancy is currently unpredictable due to the fact that the underlying pathophysiology is not fully elucidated. Regulation of apoptosis is a potential way to suppress immune activation during pregnancy. Pro-apoptotic mediators are released from the placenta and can be involved in the induction of increased T cell

apoptosis [4,5]. Circulating apoptotic bodies and micro-particles are possible mediators for these apoptotic signals during gestation [6]. In result, an increased prevalence of apoptotic (CD95+) T cells is reported in healthy pregnant compared to healthy non-pregnant women [7]. Interestingly, recent studies reported disturbance in the pro- and anti-apoptotic balance when gestation accompanied by asthma [7,8]; however this has not been studied in details.

Recent research focused on the role of anti-apoptotic Birc5 protein, also known as survivin, in physiological and pathological pregnancies. For brevity, we will henceforth refer to Birc5/survivin as survivin. Survivin is a member of the inhibitor-of-apoptosis family which inhibits the caspase-regulated apoptotic pathway [9]. It plays an essential role during foetal life by regulating normal cytotrophoblast development [10,11] and survivin is highly expressed in various malignancies [12]. Its function in adult differentiated cells is not fully known, but it may regulate activation and proliferation of T cells [13].

* Correspondence: andras.bikov@gmail.com

¹Department of Pulmonology, Semmelweis University, 1/C Dios arok, Budapest H-1125, Hungary

Full list of author information is available at the end of the article

During pregnancy survivin is produced mostly in cytotrophoblast and weakly in syncytiotrophoblast cells of the placenta [10]. It is responsible for cytotrophoblast survival by regulating cell mitosis [11]. Its role in pathological pregnancies is controversial. In hydatidiform moles and choriocarcinomas survivin levels were elevated [10,14] while in preeclampsia decreased expression has been reported [11,15]. Although no study has examined circulating survivin concentration in pregnancy, it is hypothesised that survivin levels are decreased as a consequence of up-regulated pro-apoptotic processes.

Survivin may also be involved in chronic inflammatory diseases such as bronchial asthma. Extracellular survivin promotes the differentiation of T cells toward Th2 line and enhances the production of some type 2 cytokines, including IL-4 and IL-13 [16]. The gene expression of survivin increases in ovalbumin-induced asthmatic mice [17,18] and in induced sputum samples of asthmatic patients [19]. Moreover, sputum survivin mRNA levels are related to airway eosinophilia [19]. Interestingly, certain single nucleotide polymorphisms of the survivin gene are more likely associated with asthma in women [19].

However, survivin is difficult to investigate in asthmatic pregnancy as direct airway sampling methods, such as bronchial biopsy or bronchoalveolar lavage, are invasive and cannot be performed. Similarly, placental sampling also carries risk for complications. The analysis of circulating survivin is a harmless and promising method to study survivin-related processes [20-23], especially in solid tumours and leukaemia [20]. However plasma survivin has not been studied either in pregnancy or in asthma before.

As survivin is involved in asthma and pathological pregnancy, we hypothesised that it may be altered in asthmatic gestation. To investigate this, plasma survivin levels were measured in asthmatic and healthy pregnant and non-pregnant women.

Methods

Study subjects

Twenty-eight asthmatic pregnant (AP, 31 ± 5 years), 25 asthmatic non-pregnant (ANP, 32 ± 7 years), 21 healthy pregnant (HP, 31 ± 5 years) and 29 healthy non-pregnant (HNP, 30 ± 5 years) women were enrolled. The AP group comprised volunteers in the 2nd trimester ($N = 19$, 20 ± 5 gestational weeks) or 3rd trimester ($N = 9$, 34 ± 4 gestational weeks), while the HP group consisted of participants in the 2nd trimester (23 ± 3 gestational weeks). All volunteers were Caucasian except for one AP women who had Asian origin.

Asthmatic patients were recruited at the outpatient clinic of Department of Pulmonology. Asthma was diagnosed by a respiratory medicine specialist according to the Global Initiative for Asthma (GINA) guidelines. Asthmatic patients with exacerbations within the last

6 months were not included. Nineteen ANP and fifteen AP subjects used inhaled corticosteroids regularly, while others were considered steroid-naive. The asthma was considered well-controlled or partially controlled in 13 ANP and 14 AP subjects and uncontrolled in 12 and 14 subjects, respectively.

Pregnant women were recruited at the First Department of Obstetrics and Gynecology. In all cases, the pregnancy and labour were uncomplicated and pregnant women gave birth to healthy children. Volunteers with twin pregnancies or in whom later preeclampsia developed were not studied. HNP volunteers were workers and students of Semmelweis University.

Subjects with any chronic disease, including hypertension, diabetes or malignancies were excluded. None of the participants were current or ex-smokers or had any respiratory tract infection within 4 weeks of the study.

Study design

In all subjects, venous blood was collected in EDTA-tubes. In eight 2nd-trimester asthmatic pregnant volunteers, sample collection was repeated in the 3rd trimester (in the cross-sectional analysis only the sample from the 2nd trimester was used). In asthmatic subjects, additional lung function and fractional exhaled nitric oxide (FE_{NO}) [24] measurements were performed and asthma control was evaluated with the Asthma Control Test (ACT) [25].

The study was approved by the Semmelweis University Ethics Committee (TUKEB 110/2007), and all patients gave written informed consent prior to participation in the study.

Plasma survivin measurements

Plasma was separated according to the ELISA kit guidelines and stored at -80°C until survivin measurements. Plasma survivin levels were determined by a commercially available ELISA kit (DSV00, R&D Systems, Abingdon, UK). The detection limit was 4.44 pg/ml, as it was reported by the manufacturer. The mean intra-assay coefficient for variation of duplicate samples was 22%.

Statistical analysis

We used Graphpad Prism 4.0 (GraphPad Software Inc., San Diego, CA, USA) for statistical analysis. The normality distribution of the data was assessed by Kolmogorov-Smirnov test. Plasma survivin was compared among groups with two-way ANOVA followed by Bonferroni post hoc test. Unpaired t-test was applied to compare lung function variables, steroid use and neonatal birth weight, while Mann-Whitney test was used to compare FE_{NO} and ACT levels. The relationship between survivin levels and clinical variables was analysed with Spearman tests. Pearson and Spearman tests were used to correlate clinical variables within groups. Since plasma survivin as

well as FE_{NO} levels were not normally distributed, these variables were expressed as median/range/, otherwise as mean ± SD. Samples with plasma survivin levels below the detection limit were assigned to have 0 pg/ml of survivin. $p < 0.05$ was considered significant.

The sample size was calculated to find differences in plasma survivin levels among the four groups using an effect size of 0.35 and a statistical power (1-β) of 0.80 taking into account the asymptotic relative efficiency of non-parametric tests [26].

Results

Comparison of the four groups

The two asthmatic groups (AP and ANP) were comparable in terms of lung function, inhaled corticosteroid use, FE_{NO} and asthma control (all $p > 0.05$). Similarly, there was no difference in neonatal birth weight, week of delivery or in the 0- and 5-minute Apgar scores between the AP and HP groups (all $p > 0.05$, Table 1).

Circulating survivin levels and their relationship to clinical parameters

Survivin was detectable in 61% of AP, 43% of HP, 68% of ANP and 72% of HNP women. Comparing the four groups using two-way ANOVA, significantly lower plasma survivin levels were noted in pregnancy ($p = 0.04$), however asthma had no effect ($p = 0.59$). Bonferroni post hoc test revealed that pregnancy-related differences were present only in healthy groups (1.64/0-74.9 pg/ml/ vs. 24.6 /0-333.3/ pg/ml, $p = 0.01$, HP vs. HNP, respectively), while there was no difference when the asthmatic non-pregnant (10.5 /0-215.4/ pg/ml) and asthmatic pregnant (13.9/0-364.1/) patients were compared ($p = 0.64$). If the 3 outliers in the AP group were excluded the difference between the 4 groups was still significant ($p < 0.01$). Comparing asthmatic patients to the corresponding non-asthmatic

subjects no difference was found either between pregnant ($p = 0.30$) or non-pregnant ($p = 0.23$) groups (Figure 1). When AP patients only in the 2nd trimester were compared to HP volunteers the difference was still not significant ($p = 0.25$).

There was no difference between AP subjects in the 2nd (17.8 /0-364.1/ pg/ml) and 3rd trimester (0 /0-185.0/ pg/ml, $p = 0.61$). Nor were the survivin levels in the same individual (8 AP subjects) different between the two time points (13.1 /0-31.6/ pg/ml vs. 6.6 /0-121.0/ pg/ml, 2nd vs. 3rd trimester, respectively, $p = 0.79$, Figure 2). This indicates that differences in gestational age between the AP and HP groups did not bias the results on survivin. Comparing survivin levels depending on newborn gender there was no difference either in AP ($p = 0.47$) or HP ($p = 0.45$) groups.

The relationships between plasma survivin levels and lung function variables, FE_{NO} or asthma control in the AP and ANP groups were not significant (all $p > 0.05$). Nor was there any relationship between plasma survivin and gestational weeks or neonatal birth weight in either of the AP and HP groups (all $p > 0.05$). Comparing asthmatic patients using inhaled corticosteroids (ICS) with steroid-naive subjects, no difference was observed in plasma survivin either in the ANP ($p = 0.80$) or AP groups ($p = 0.58$). Similarly, there was no relationship between ICS dose and plasma survivin levels either in ANP ($p = 0.19$) or AP ($p = 0.69$) subjects.

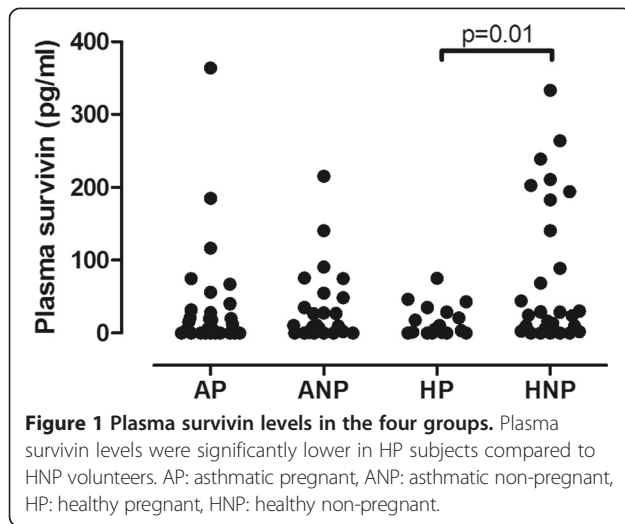
Relationship between clinical variables

In the ANP group a significant relationship was found between FEV₁ and ACT ($r = 0.43$, $p = 0.03$) and there was a statistical tendency for inverse correlation between FEV₁ and FE_{NO} levels ($r = -0.44$, $p = 0.06$). Interestingly, these correlations were not present in AP subjects either when the 2nd and 3rd trimester groups were analysed together or separately.

Table 1 Clinical characteristics of study subjects

	AP N = 28	ANP N = 25	HP N = 21	HNP N = 29	P value
FEV ₁ ; L	2.9 ± 0.4	2.8 ± 0.7			0.54
(% pred)	(90 ± 11)	(88 ± 18)	ND	ND	(0.66)
FVC; L	3.6 ± 0.5	3.7 ± 0.8			0.85
(% pred)	(99 ± 13)	(100 ± 15)	ND	ND	(0.68)
FE _{NO} ; ppb	19 (8–115)	19 (5–82)	ND	ND	0.62
ACT	20 (8–25)	20 (9–25)	ND	ND	0.76
ICS; BDP eq.	200 (0–2000)	400 (0–1000)	NA	NA	0.25
Neonatal birth weight; g	3548 ± 714	NA	3442 ± 320	NA	0.59
Apgar; 1 and 5 minutes	9 and 10	NA	9 and 10	NA	0.27 and 0.95
Week of delivery	39 (36–42)	NA	39 (36–41)	NA	0.44

AP: asthmatic pregnant, ANP: asthmatic non-pregnant, HP: healthy pregnant, HNP: healthy non-pregnant, ACT: asthma control test, BDP eq.: beclomethasone dipropionate equivalent, FE_{NO}: fractional exhaled nitric oxide, FEV₁: forced expiratory volume in one second, FVC: forced vital capacity, ICS: inhaled corticosteroid, NA: not applicable, ND: not determined, ppb: particles per billion. Data are expressed as mean ± SD or median (range).



Discussion

In the current study we investigated plasma survivin levels in asthma, together with asthmatic and healthy pregnancies. We found that circulating survivin is decreased during gestation, which was blunted in asthmatic pregnancy.

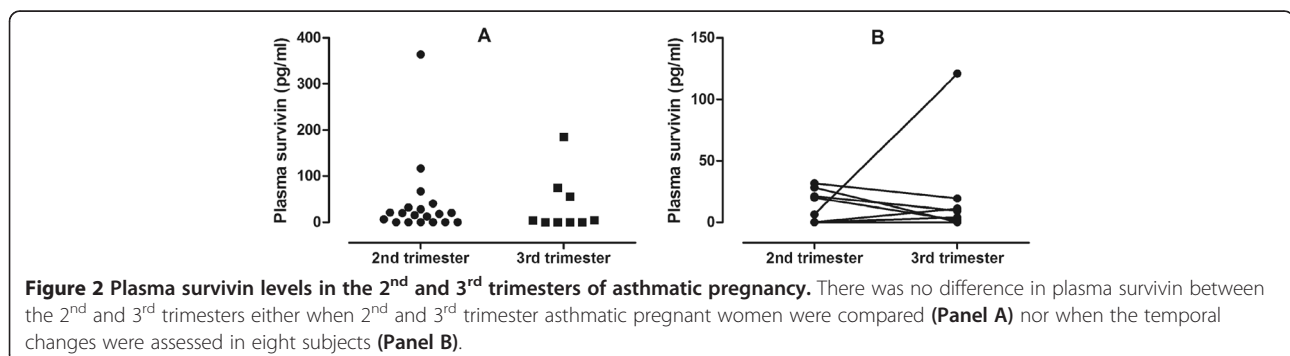
This is the first study analysing circulating survivin levels in pregnancy, however the intracellular expression during gestation has already been investigated in physiological and pathological circumstances. It is known that this molecule is produced by the placenta and has an important role in the normal cytotrophoblast development [10,11]. Its expression is tightly regulated as both increased and reduced productions are associated with pathological pregnancies [10,11,14,15].

The source and function of extracellular survivin in pregnancy is not known. It may originate from dead cells, but survivin can also be actively released by living cells [27]. In malignancies, extracellular survivin is taken up by the surrounding cancer cells inhibiting their apoptosis, accelerating their proliferation and increasing their invasive potential [27]. A recent study described that survivin produced by cancer cells also inhibits T cell

activation and proliferation [16]. Hence, decreased survivin levels in pregnancy may be associated with enhanced T cell activation. Supporting this, we have previously reported that non-asthmatic pregnancy is associated with activation and apoptosis of T cells [7]. Survivin affects lymphocyte subtypes in different ways. It decreases the number and suppresses the function of CD8+ T cells, skewing immunity towards the Th2 direction, but not altering the regulatory T cell and Th17 ratios [16]. In addition, IFN- γ and IL-2 levels are decreased while IL-4 and IL-13 concentrations are increased in the presence of survivin [16]. It is known that healthy pregnancy is associated with altered T cell balance [28] and cytokine profiles [29] with elevated proportions of CD8+ cells [28] and decreased levels of IL-4 [29].

Another possible reason for low extracellular survivin in pregnancy might be the reduced production of vascular endothelial growth factor (VEGF) [30]. It is known that the expression of survivin is induced by VEGF [31] which is supported by a significant relationship between plasma survivin and VEGF levels [21]. Finally, it is known that survivin is down-regulated by progesterone [32] the level of which is highly elevated during gestation. As we only investigated pregnant subjects in the 2nd and 3rd trimesters, we do not know if extracellular survivin is equally low during the whole course of pregnancy. Only one study measured placental survivin mRNA, showing reducing levels throughout the pregnancy [15]. We did not find significant differences in survivin levels between the 2nd- and 3rd-trimester pregnant asthmatics, however this analysis was poorly powered in the current study and we cannot rule out the possibility that survivin levels may change in non-asthmatic pregnant subjects.

Asthma may modulate pregnancy-related immune responses [28,33]. For instance, CD8+ T cell prevalence is decreased [28], while IL-4 and IFN- γ levels are elevated [33] in asthmatic gestation. The absence of a physiological decrease in survivin levels in asthmatic pregnancy may contribute to these immunological changes. Nonetheless, IFN- γ , known to be increased in asthmatic



pregnancy [33] may up-regulate survivin expression [13] contributing to its blunted decrease seen in the current study.

In addition, various studies suggest that asthma also suppresses the anti-apoptotic mechanism seen in physiological pregnancy. We have previously described that lymphocyte apoptosis is enhanced in healthy pregnancy; however this effect is limited in asthmatic pregnant women [7]. Similarly, another anti-apoptotic agent [34], heat shock protein 70 was also found to be decreased in normal pregnancy [35], but not in asthmatic pregnant women [8]. Our present results are consistent with these previous findings.

Recent studies supported the role of survivin in the pathomechanism of asthma [17-19]. We could not find any differences between asthmatic and non-asthmatic subjects either when the pregnant or non-pregnant women were compared. Similarly, there was no correlation with any of the asthma variables. This might suggest that survivin-related asthmatic processes are localised in the lungs, as in the previous human study, only airway samples were analysed [19]. In the present study, asthmatic subjects with recent exacerbation were excluded, and asthma was considered relatively stable in participants. Despite the fact that there was no association between survivin levels and clinical variables of asthma, we cannot exclude the possibility that heightened disease activity (i.e. during exacerbation) might be related with increased systemic survivin. Of note, increased survivin in induced sputum samples of asthmatic patients was noted even in stable subjects [19].

Only a few studies have examined plasma survivin to date in various diseases including solid tumours, leukaemia, rheumatoid arthritis and HCV infection [20-23]. In overall, the median values were very close to the lower limit of detection with around 30% of the samples below the limit of detection which is in line with the observations of the current study. Survivin was even more poorly detectable in pregnancy which further confirms that this molecule is decreased during gestation, but unfortunately this also limits the statistical power of our conclusions. Previous studies measured survivin in plasma samples with ELISA which is a more feasible method to analyse survivin in <100 pg/ml concentration range than Western blot which has a detection limit around 100 pg/ml in plasma samples. Two studies used the same commercially available ELISA kit as in our study [21,23], but unfortunately neither of them reported intra-assay variability. The relatively poor analytical repeatability of this analytical method and the high proportion of samples with survivin concentration below the detection limit may not allow to draw final conclusions from some analyses done on small number of samples (i.e. the effect of gestational age or clinical outcomes of

asthma or gestation). Therefore, the results on these statistics should be interpreted carefully. Of note, further studies are warranted to optimise the medium (serum, EDTA or citrate plasma) for survivin measurements, as the healthy values tended to be higher in a previous study using citrate tubes than in our results measured in samples collected in EDTA tubes [23].

Conclusions

In summary, for the first time, we reported significantly lower levels of plasma survivin in physiological but not in asthmatic gestation. Further studies are warranted to fully investigate the influence of survivin on apoptotic and immunological processes in pregnancy.

Abbreviations

ACT: Asthma control test; AP: Asthmatic pregnant; ANP: Asthmatic non-pregnant; BDP eq.: Beclomethasone dipropionate equivalent; FENO: Fractional exhaled nitric oxide; FEV1: Forced expiratory volume in one second; FVC: Forced vital capacity; HP: Healthy pregnant; HNP: Healthy non-pregnant; ICS: Inhaled corticosteroid; LD: Limit of detection; SD: Standard deviation; VEGF: Vascular endothelial growth factor.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AB carried out survivin measurements and drafted the manuscript. RB, NE and AB contributed to recruiting and clinically characterizing patients and participated in sample collection. GL, JR and IH contributed to the design and coordination of the study and provided facilities for the measurements. LT conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The work was supported by the Hungarian Scientific Research Fund (OTKA 68808) and Hungarian Respiratory Society (grants to Andras Bikov and Renata Bocskai). The authors are extremely grateful to Dr. Sophia Lazar for English corrections.

Author details

¹Department of Pulmonology, Semmelweis University, 1/C Dios arok, Budapest H-1125, Hungary. ²First Department of Obstetrics and Gynecology, Semmelweis University, 27 Baross utca, Budapest H-1085, Hungary.

Received: 27 May 2014 Accepted: 18 September 2014

Published: 23 September 2014

References

1. Tamasi L, Somoskovi A, Muller V, Bartfai Z, Acs N, Puho E, Czeizel AE: A population-based case-control study on the effect of bronchial asthma during pregnancy for congenital abnormalities of the offspring. *J Asthma* 2006, **43**:81-86.
2. Dombrowski MP: Asthma and pregnancy. *Obstet Gynecol* 2006, **108**:667-681.
3. Maselli DJ, Adams SG, Peters JI, Levine SM: Management of asthma during pregnancy. *Thorax* 2013, **7**:87-100.
4. Frangsmyr L, Baranov V, Nagaeva O, Stendahl U, Kjellberg L, Mincheva-Nilsson L: Cytoplasmic microvesicular form of Fas ligand in human early placenta: switching the tissue immune privilege hypothesis from cellular to vesicular level. *Mol Hum Reprod* 2005, **11**:35-41.
5. Taylor DD, Sullivan SA, Eblen AC, Gerceel-Taylor C: Modulation of T-cell CD3-zeta chain expression during normal pregnancy. *J Reprod Immunol* 2002, **54**:15-31.
6. Redman CW, Sargent IL: Circulating microparticles in normal pregnancy and pre-eclampsia. *Placenta* 2008, **29**(A):S73-S77.

7. Bohacs A, Pallinger E, Tamasi L, Rigo J Jr, Komlosi Z, Muller V, Dong Y, Magyar P, Falus A, Losonczy G: **Surface markers of lymphocyte activation in pregnant asthmatics.** *Inflamm Res* 2010, **59**:63–70.
8. Tamasi L, Bohacs A, Tamasi V, Stenczer B, Prohaszka Z, Rigo J Jr, Losonczy G, Molvarec A: **Increased circulating heat shock protein 70 levels in pregnant asthmatics.** *Cell Stress Chaperones* 2010, **15**:295–300.
9. Cheung CH, Huang CC, Tsai FY, Lee JY, Cheng SM, Chang YC, Huang YC, Chen SH, Chang JY: **Survivin - biology and potential as a therapeutic target in oncology.** *Onco Targets Ther* 2013, **6**:1453–1462.
10. Shiozaki A, Kataoka K, Fujimura M, Yuki H, Sakai M, Saito S: **Survivin inhibits apoptosis in cytotrophoblasts.** *Placenta* 2003, **24**:65–76.
11. Muschol-Steinmetz C, Friemel A, Kreis NN, Reinhard J, Yuan J, Louwen F: **Function of survivin in trophoblastic cells of the placenta.** *PLoS One* 2013, **8**:e73337.
12. Adida C, Berrebi D, Peuchmaur M, Reyes-Mugica M, Altieri DC: **Anti-apoptosis gene, survivin, and prognosis of neuroblastoma.** *Lancet* 1998, **351**:882–883.
13. Zimmerman M, Yang D, Hu X, Liu F, Singh N, Browning D, Ganapathy V, Chandler P, Choubey D, Abrams SI, Liu K: **IFN-gamma upregulates survivin and lfi202 expression to induce survival and proliferation of tumor-specific T cells.** *PLoS One* 2010, **5**:e14076.
14. Lehner R, Bobak J, Kim NW, Shroyer AL, Shroyer KR: **Localization of telomerase hTERT protein and survivin in placenta: relation to placental development and hydatidiform mole.** *Obstet Gynecol* 2001, **97**:965–970.
15. Li CF, Gou WL, Li XL, Wang SL, Yang T, Chen Q: **Reduced expression of survivin, the inhibitor of apoptosis protein correlates with severity of preeclampsia.** *Placenta* 2012, **33**:47–51.
16. Jutzy JM, Khan S, Asuncion-Valenzuela MM, Milford TA, Payne KJ, Wall NR: **Tumor-released survivin induces a type-2 t cell response and decreases cytotoxic T cell function, in vitro.** *Cancer Microenviron* 2013, **6**:57–68.
17. Ungvari I, Hullam G, Antal P, Kiszal PS, Gezsi A, Hadadi E, Virag V, Hajos G, Millinghoff A, Nagy A, Kiss A, Semsei AF, Temesi G, Melegh B, Kisfali P, Szell M, Bikov A, Galffy G, Tamasi L, Falus A, Szalai C: **Evaluation of a partial genome screening of two asthma susceptibility regions using bayesian network based bayesian multilevel analysis of relevance.** *PLoS One* 2012, **7**:e33573.
18. Tumes DJ, Connolly A, Dent LA: **Expression of survivin in lung eosinophils is associated with pathology in a mouse model of allergic asthma.** *Int Immunol* 2009, **21**:633–644.
19. Ungvari I, Hadadi E, Virag V, Bikov A, Nagy A, Semsei AF, Galffy G, Tamasi L, Horvath I, Szalai C: **Implication of BIRC5 in asthma pathogenesis.** *Int Immunol* 2012, **24**:293–301.
20. Sugahara K, Uemura A, Harasawa H, Nagai H, Hirakata Y, Tomonaga M, Murata K, Sohda H, Nakagoe T, Shibasaki S, Yamada Y, Kamihira S: **Clinical relevance of survivin as a biomarker in neoplasms, especially in adult T-cell leukemias and acute leukemias.** *Int J Hematol* 2004, **80**:52–58.
21. Yang M, Liu Y, Lu S, Wang Z, Wang R, Zi Y, Li J: **Analysis of the expression levels of survivin and VEGF in patients with acute lymphoblastic leukemia.** *Exp Ther Med* 2013, **5**:305–307.
22. El-Attar HA, Kandil MH, El-Kerm YM, El-Ghandour MK: **Comparison of serum survivin and alpha fetoprotein in Egyptian patients with hepatocellular carcinoma associated with hepatitis C viral infection.** *Asian Pac J Cancer Prev* 2010, **11**:897–903.
23. Bokarewa M, Lindblad S, Bokarew D, Tarkowski A: **Balance between survivin, a key member of the apoptosis inhibitor family, and its specific antibodies determines erosivity in rheumatoid arthritis.** *Arthritis Res Ther* 2005, **7**:R349–R358.
24. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide, 2005. *Am J Respir Crit Care Med* 2005, **171**:912–930.
25. Nathan RA, Sorkness CA, Kosinski M, Schatz M, Li JT, Marcus P, Murray JJ, Pendergraft TB: **Development of the asthma control test: a survey for assessing asthma control.** *J Allergy Clin Immunol* 2004, **113**:59–65.
26. Faul F, Erdfelder E, Buchner A, Lang AG: **Statistical power analyses using G*Power 3.1: tests for correlation and regression analyses.** *Behav Res Methods* 2009, **41**:1149–1160.
27. Khan S, Aspe JR, Asumen MG, Almaguel F, Odumoso O, Acevedo-Martinez S, De Leon M, Langridge WH, Wall NR: **Extracellular, cell-permeable survivin inhibits apoptosis while promoting proliferative and metastatic potential.** *Br J Cancer* 2009, **100**:1073–1086.
28. Toldi G, Molvarec A, Stenczer B, Muller V, Eszes N, Bohacs A, Bikov A, Rigo J Jr, Vasarhelyi B, Losonczy G, Tamasi L: **Peripheral T(h)1/T(h)2/T(h)17/regulatory T-cell balance in asthmatic pregnancy.** *Int Immunol* 2011, **23**:669–677.
29. Molvarec A, Szarka A, Walentin S, Beko G, Karadi I, Prohaszka Z, Rigo J Jr: **Serum leptin levels in relation to circulating cytokines, chemokines, adhesion molecules and angiogenic factors in normal pregnancy and preeclampsia.** *Reprod Biol Endocrinol* 2011, **9**:124.
30. Bikov A, Bohacs A, Eszes N, Weiszhar Z, Ivancso I, Muller V, Rigo J Jr, Losonczy G, Tamasi L, Horvath I: **Circulating and exhaled vascular endothelial growth factor in asthmatic pregnancy.** *Biomarkers* 2012, **17**:648–654.
31. Beierle EA, Nagaram A, Dai W, Iyengar M, Chen MK: **VEGF-mediated survivin expression in neuroblastoma cells.** *J Surg Res* 2005, **127**:21–28.
32. Formby B, Wiley TS: **Bcl-2, survivin and variant CD44 v7-v10 are downregulated and p53 is upregulated in breast cancer cells by progesterone: inhibition of cell growth and induction of apoptosis.** *Mol Cell Biochem* 1999, **202**:53–61.
33. Tamasi L, Bohacs A, Pallinger E, Falus A, Rigo J Jr, Muller V, Komlosi Z, Magyar P, Losonczy G: **Increased interferon-gamma- and interleukin-4-synthesizing subsets of circulating T lymphocytes in pregnant asthmatics.** *Clin Exp Allergy* 2005, **35**:1197–1203.
34. Beere HM, Wolf BB, Cain K, Mosser DD, Mahboubi A, Kuwana T, Tailor P, Morimoto RI, Cohen GM, Green DR: **Heat-shock protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome.** *Nat Cell Biol* 2000, **2**:469–475.
35. Molvarec A, Tamasi L, Losonczy G, Madach K, Prohaszka Z, Rigo J Jr: **Circulating heat shock protein 70 (HSPA1A) in normal and pathological pregnancies.** *Cell Stress Chaperones* 2010, **15**:237–247.

doi:10.1186/1477-7827-12-93

Cite this article as: Bikov et al.: Circulating survivin levels in healthy and asthmatic pregnancy. *Reproductive Biology and Endocrinology* 2014 **12**:93.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

