Human and Experimental Toxicology Online: pp. 1-11 (2013)

http://het.sagepub.com/content/early/2013/05/06/0960327113480972

Protective effect of CV247 against cisplatin nephrotoxicity in rats

Csaba Máthé<sup>1</sup>, Gábor Szénási<sup>2</sup>, Andor Sebestény<sup>3</sup>, Anna Blázovics<sup>4</sup>, Klára Szentmihályi<sup>5</sup>,

Péter Hamar<sup>2</sup>, and Mihály Albert<sup>6</sup>

<sup>1</sup>Department of Pulmonology, Semmelweis University, Budapest, Hungary;

<sup>2</sup>Institute of Pathophysiology, Semmelweis University, Budapest, Hungary;

<sup>3</sup>Laboratory Animal Science Unit, Faculty of Veterinary Science, Szent István University,

Budapest, Hungary;

<sup>4</sup>Department of Pharmacognosy, Semmelweis University, Budapest, Hungary;

<sup>5</sup>Institute of Materials and Environmental Chemistry Research Centre for Natural Sciences,

Budapest, Hungary;

<sup>6</sup>Vetmed Laboratory Ltd. Budapest, Hungary;

\* Corresponding author: Gábor Szénási, PhD

Address: Institute of Pathophysiology, Semmelweis University, 1089 Nagyvárad tér 4,

Budapest, Hungary.

Tel: +36 20 8259229

E-mail address: szenasi.gabor@med.semmelweis-univ.hu

Conflict of Interest: None

# **Abbreviations**

Crea serum creatinine

BUN blood urea nitrogen

COX-2 prostaglandin-endoperoxide synthase 2 (cyclooxygenase-2)

C control

CV CV247

CDDP cis-diammine-dichloroplatinum; trade name: Cisplatin

ICP-OES inductively coupled plasma optical emission spectrometry

#### **Abstract**

CV247, an aqueous mixture of copper and manganese gluconates, vitamin C and sodium salicylate, increased the anti-tumour effects of cisplatin *in vitro*. We hypothesized that the antioxidant and cyclooxygenase (COX-2) inhibitory components of CV247 can protect the kidneys from cisplatin nephrotoxicity in rats.

Cisplatin (6.5 mg/kg, ip.) slightly elevated serum creatinine (Crea) and blood urea nitrogen (BUN) 12 days after treatment. Kidney histology demonstrated extensive tubular epithelial damage, and COX-2 immunoreactivity increased 14 days after treatment. Cisplatin increased renal platinum (Pt) but decreased iron (Fe), copper (Cu), manganese (Mn), molybdenum (Mo) and zinc (Zn) concentrations, and increased plasma Fe and Cu concentrations. Cisplatin elevated plasma free radical concentration. Treatment with CV247 alone for 14 days (twice 3 ml/kg/day p.o.) did not influence these parameters.

Chronic CV247 administration after cisplatin reduced renal histological damage and almost significantly decreased COX-2 immunoreactivity, while failed to improve Crea and BUN. Blood free radical concentration was decreased, i.e. CV247 improved redox homeostasis. CV247 restored plasma Fe and renal Fe, Mo and Zn, while decreased renal Pt and elevated Cu and Mn concentrations.

Besides the known synergistic anti-tumour effects with cisplatin, CV247 partially protected the kidneys from cisplatin nephrotoxicity probably through its antioxidant effect.

Keywords: antioxidant, cisplatin, COX-2, manganese, nephrotoxicity, salicylate

#### Introduction

Cisplatin is a highly effective chemotherapeutic agent used for the treatment of various malignancies. High-dose cisplatin-based combination chemotherapy regimens are used as first-line treatment of small-cell and non-small cell lung cancers. However, the use of cisplatin is limited by serious side effects. Despite the use of different hydration protocols allowing dose escalation to therapeutic levels, nephrotoxicity is the main dose-limiting side effect of cisplatin. About 20% of acute renal failure cases was due to cisplatin among hospitalised patients. High-dose cisplatin-induced nephrotoxicity (>25% decrease in eGFR) was diagnosed in 29 % of patients, following a single dose of cisplatin, and temporary elevation of serum creatinine concentration above the upper normal limit was observed in 41% of 400 cisplatin-treated patients with different solid tumours. Comorbidities in lung cancer patients greatly increased the incidence of cisplatin-induced nephrotoxicity from 7.5% without co-morbidities to 20.9% with concurrent hypertension with or without ischemic heart disease, and to 30.8% with diabetes mellitus and ischaemic heart disease. Due to the superior efficacy of cisplatin against a variety of human carcinomas, intensive efforts have been undertaken to weaken the side effects especially the nephrotoxic effect of cisplatin.

During treatment, cisplatin accumulates in the kidney and thus, by far the highest cisplatin concentration is measured in the kidney after treatment. Dopper transporter 1 (Ctr1) and the organic cation transporter 2 (OCT2) are critically involved in cisplatin uptake into renal tubular epithelial cells consequently determining nephrotoxicity DNA damage are also implicated in the mechanisms of cisplatin-induced nephrotoxicity. The importance of oxidative stress has been highlighted by numerous studies demonstrating, that antioxidant agents ameliorate cisplatin nephrotoxicity in experimental animals. The organic thiophosphate amifostine is an FDA approved protective agent against cisplatin nephrotoxicity although its efficacy could not be

unequivocally demonstrated in animal experiments. Alkaline phosphatase converts amifostine to a free thiol compound, which is citoprotective by binding free radicals. Acute or chronic treatment with vitamin C produced encouraging results in rat experiments. Coadministration of Vitamin E and selenium were also beneficial. Acetylsalicylic acid and sodium salicylate were also protective. Acetylsalicylic acid and sodium salicylate were also protective.

CV247 is composed of manganese and copper gluconates, sodium salicylate and ascorbic acid (exact composition is given in the methods section), which are known to have antioxidant (ascorbic acid, manganese and copper), and TNFα inhibitor (sodium salicylate) effects. CV247 was shown to decrease the viability of six cancer cell lines in culture and to augment the cytotoxic effect of cisplatin against human breast and especially colon carcinoma. As some constituents of CV247 protected against kidney injury as shown above, it may also alleviate the cisplatin-induced nephrotoxicity by synergistically acting at several target sites. Therefore, the aim of this study was to investigate the net effects of CV247 on renal function, antioxidant status and kidney histology after cisplatin treatment.

#### Materials and methods

#### Animals

The study was conducted on 40 male, 8-week old Wistar rats weighing 175-190 g. The animals were randomly divided into 4 groups (n=10/group). They were kept individually under standard conventional conditions according to European Council Directive 123. The study conformed to the Declaration of Helsinki guidelines and was approved by the local Animal Ethic Committee.

### Test materials

Cisplatin (10 mg in 20 ml) was obtained from TEVA, Israel. The composition of CV247 (Pharmaserve Ltd, Manchester UK) was the following: 40 mg ascorbic acid, 2 mg manganese gluconate (USP), 2 mg copper gluconate, 35 mg sodium salicylate per millilitre solution (www.ivymedical.com). Methyl cellulose mucilage (Dow Chemicals) was prepared in distilled water (1 %).

### *Protocol of the study*

The control group (C) received 1% methyl cellulose at 10 ml/kg body weight, p.o. by gastric gavage twice daily for 14 days. Another group of rats received CV247 at 3 ml/kg body weight, p.o. twice daily for 14 days (CV). The dose given was 2x120 mg/kg/day vitamin C, 2x105 mg/kg/day sodium salicylate, 2x6 mg/kg/day copper gluconate and 2x6 mg/kg/day manganese gluconate. Two groups were intraperitoneally injected with a single dose of cisplatin (CDDP) at 6.5 mg/kg body weight. CDDP was suspended in 10 ml/kg 1% methyl cellulose. One of the groups injected with CDDP was subsequently treated with vehicle (C) or CV at 3 ml/kg body weight, p.o. twice daily for 14 days (CDDP+CV). All rats were weighed and food and water consumptions were also measured daily. On day 12 1.5 ml blood samples were taken from all rats by retro orbital puncture under isoflurane anaesthesia after a 20-hour food deprivation. The blood was anticoagulated with citrate and centrifuged twice at 2500

rpm for 10 min at +4 °C to obtain plasma. Plasma creatinine (Crea) and blood urea nitrogen (BUN) were determined from the plasma by colorimetric tests using commercially available kits. Rats were terminally anaesthetised with pentobarbitone on day 14. Blood was collected by aortic puncture and the kidneys were removed and weighed.

Induced chemiluminescence in the plasma

A small volume of plasma samples (50-100 µl) were assayed with a H<sub>2</sub>O<sub>2</sub>/OH–microperoxidase-luminol system for 30 sec as described previously.<sup>42</sup> Chemiluminescence was detected in a Berthold Lumat 9501 luminometer (Berthold GmbH, Germany).

Metal contents in the plasma and kidney

After digestion of the samples in nitric acid (5 ml 65%) and hydrogen peroxide (2 ml 30%), an inductively coupled plasma optical emission spectrometric (ICP-OES) method was used for measuring metal content<sup>43</sup> in a Spectro Genesis ICP equipment (Kleve, Germany). For the standardization of equipment and measurements of elements Spectro multielement and Spectrum 3D standards were used. A computer guided TraceLab 50 type polarographic-voltammetric analyser was used for the voltammetric determination of selenium at -550 mV.<sup>44</sup> *Histology and immunohistochemistry* 

The kidneys were fixed in 8% buffered formalin (pH 7.4) and paraffin sections were prepared and stained with haematoxylin–eosin. Renal histological changes were blindly evaluated using a 5-grade severity scale (0 = no change; 1 = minimal changes; 2 = mild changes; 3 = moderate changes; 4 = severe changes). The cyclooxygenase-2 (COX-2) immunohistochemistry was done using a mouse monoclonal COX-2 primary antibody (Novocastra, UK) at 1:100 dilution. The secondary antibody was a peroxidase-conjugated mouse/rabbit polymer (Dako Real<sup>TM</sup> Envision<sup>TM</sup> /HRP, Rabbit/Mouse). Diaminobenzidine was used for visualisation.

Statistical analysis

Means±SD are given throughout. The statistical comparisons were performed by two-way repeated measures ANOVA with Bonferroni post hoc test or Mann-Whitney U test using GraphPad Prism 5 for Windows, or by two-way ANOVA using the SPSS 17 for Windows, when appropriate. The level of significance was set at p<0.05.

#### **Results**

### **Bodyweight**

Body weight of rats steadily increased in group C from 171±8 g to 234±15 g over the 14 days of the study with a drop on day 11 after the overnight food deprivation (Fig. 1). In comparison to the baseline value, CDDP caused a 5.5 % peak body weight loss (p<0.001) from 167±6 g to 158±9 g on day 3 after treatment. Thereafter, body weight gain returned to a rate similar to that seen in group C. CV treatment did not influence body weight in comparison to groups C and CDDP, respectively. Consequently, on the last day of the study, body weight of rats treated with CDDP and CDDP+CV was 12-15 % lower (p<0.001) than body weight of rats in groups C and CV.

### Food and water consumptions

CV consistently increased water consumption in comparison to that in group C (**Fig. 1**), which was statistically significant on days 8, 9 and 11 (p<0.05, all). Cisplatin caused a short, non-significant decrease in water consumption on day 2 after its administration. Thereafter, from day 4 rats in the CDDP and CDDP+CV groups drank significantly more water than rats in group C. Co-administration of CV to cisplatin did not alter water consumption in comparison to the group treated with cisplatin only. Neither CDDP nor CV alone or in combination altered food consumption (data not shown).

### Clinical chemistry

Crea and BUN values were within physiological limits in groups C and CV (Crea: 17.0-22.5 µmol/l; BUN: 6.63- 10.48 mmol/l). CDDP increased both Crea and BUN concentrations at day 12 after its administration, while CV did not alter these effects of CDDP on renal function (**Fig. 2**).

Plasma reactive oxidant levels increased 14 days after CDDP administration, as measured by chemiluminescence. CV did not alter chemiluminescence in comparison to group C while CV attenuated the CDDP-induced elevation in plasma reactive oxidant levels (**Fig. 3**).

Metal concentrations in the plasma and kidneys

Kidney Cu, Fe, Mn, Mo and Zn concentrations were lower in the kidney 14 days after treatment with CDDP, while Co and Se concentrations did not change (p<0.05). Treatment with CV increased Mo concentrations in the kidney while it did not change other element concentrations (**Table 1**). Co-administration of CV with CDDP restored renal Fe and Zn concentrations to control levels, and also increased renal Cu and Mn concentrations significantly, although Cu and Mn remained below the control levels. The effect of CDDP on renal Mo concentration was restored by CV. Surprisingly, kidney Pt concentration was 3  $\mu$ g/g in the group treated with CDDP. CV strikingly reduced kidney Pt concentration by 30 % (p<0.05).

Plasma concentrations of Pt and Se were undetectable in untreated control animals. CDDP increased plasma Co and Fe concentrations (**Table 2**). Coadministration of CV with CDDP restored plasma Fe concentrations while CV did not alter the effect of CDDP on plasma Co concentrations.

Kidney histology and immunohistochemistry

No histological changes were seen in the kidneys in groups C and CV. Varying degrees of pathological changes were found in the kidneys of CDDP and CDDP+CV groups. Renal tubular epithelial cell atrophy presented as cystic dilatations of the tubular lumina, in which accumulation of desquamated tubular epithelial cells were present as hialynacous material. Many tubular epithelial cells appeared apoptotic or necrotic. In addition, in the damaged tubular epithelium atypical, regenerating cells were visible. Tubulointerstitial inflammation presented as lymphocytic and macrophage infiltration in the interstitial space, accompanied

by interstitial fibrosis appearing as multiple focal presence of fibroblasts (**Fig. 4**). Blind assessment demonstrated a reduction in the mean score severity of histological kidney injury from  $3.67\pm0.50$  in group CDDP to  $2.67\pm0.71$  in group CDDP+CV (p<0.01). Immunohistochemistry revealed a moderate degree of focal COX-2 activity in the cytoplasm of the tubular epithelium, in the interstitial space and in the walls of major blood vessels (score, control:  $1.20\pm0.42$  and CV:  $1.0\pm0.0$ ). Blind assessment of COX-2 activity revealed that COX-2 immunoreactivity markedly increased in the groups treated with CDDP and CDDP+CV. Treatment with CV did not alter COX-2 immunoreactivity in comparison to that in group C but it almost significantly  $(3.00\pm0.71 \text{ vs. } 2.44\pm0.53, \text{p=0.097}, \text{Mann Whitney test})$  decreased the changes caused by CDDP (**Fig. 5**).

#### **Discussion**

In the present study we demonstrated, that CV247, a potent enhancer of the anti-neoplastic effects of cisplatin, effectively protected the kidney from cisplatin toxicity as demonstrated by renal histology and restoration of redox and trace metal homestasis. However, slight renal retention of Crea and BUN 14 days after CDDP injection was not prevented by CV247.

In group CDDP, histological damage was clearly present at day 14 after cisplatin injection, and immunohistochemistry revealed marked Cox-2 activation in the kidney, as well as an increase in plasma reactive oxidant levels, detected by chemiluminescence. Cisplatin decreased copper, manganese and zinc concentrations in the kidney, which minerals are essential cofactors of several enzymes. Chronic treatment of rats with CV247 offered protection against cisplatin-induced nephrotoxicity demonstrated by attenuation of histological injury, restored plasma reactive oxidant levels and kidney copper content, and improved kidney manganese, selenium and zinc contents at 14 days after cisplatin administration. However, slight plasma creatinine and BUN retention still present 14 days after CDDP injection and renal inflammation, as revealed by cox-2 immunohistochemistry, were not influenced by CV247.

It has been reported that CDDP accumulated in the kidney<sup>45</sup> and renal Pt content decreased very slowly after cisplatin administration.<sup>46</sup> It was an important observation in our study that chronic treatment with CV247 significantly decreased kidney platinum content by day 14 in comparison to the cisplatin alone group. Since administration of CV247 did not precede that of cisplatin, it can be excluded that CV247 interfered with renal cisplatin uptake. Therefore it seems likely that long-term administration of CV247 accelerated elimination of Pt from the kidney. This observation suggests that kidneys of CV247-treated rats recovered faster from the cisplatin-induced nephrotoxicity.

Most importantly, blind assessment of tissue pathology demonstrated that CV247 reduced the severity of renal histological injury. This observation seems to be in harmony with attenuation or full reversal of the cisplatin-induced decreases in trace mineral content of the kidney. These changes are compatible with the assumption that concentration of all those enzymes increased in the kidney, which use these minerals as cofactors for achieving their full activity. All these changes seem to suggest that manganese and copper constituents of CV247 contributed to improve the biochemical machinery of the kidney. Although, only manganese and copper were supplemented, it is well-known that trace mineral metabolism is subject to mutual synergisms and antagonisms.<sup>47</sup> Therefore, administration of one or few trace minerals may also consequently alter the concentrations of other minerals in the kidney.

A component of CV247, sodium salicylate inhibits the activity of cyclooxygenase (COX-1 and COX-2) isoenzymes<sup>48</sup>. COX-2 inhibition is anti-inflammatory and can be a renoprotective strategy as a highly selective COX-2 inhibitor (SC-58236) reduced urinary excretions of TGF-β, TNF-α, albumin, PGE2, 6-ketoPGF1α and TxB2 in streptozotocin-diabetic rats.<sup>49</sup> Our immunohistochemistry findings demonstrated that the cisplatin-induced elevation in COX-2 expression was almost significantly decreased by chronic treatment with CV247. In addition COX-2 enzyme activity may have been reduced by CV247 as sodium salicylate inhibits COX-2.<sup>48</sup>

The protective effect of sodium salicylate could be attributed to suppression of TNFα production as well.<sup>37</sup> Treatment with acetyl salicylic acid or sodium salicylate for 4-5 days decreased plasma BUN and creatinine at a daily dose similar to that given by us<sup>38</sup>, restored the renal concentration of superoxide dismutase, and decreased oxidative stress as shown by renal malondialdehyde concentration in cisplatin nephrotoxicity.<sup>36</sup> Such effects of sodium salicylate may help explain why CV247 accelerated histological recovery of the kidney from cisplatin nephrotoxicity in our study.

Even a 10 min pretreatment with various doses of vitamin C restored cisplatin-induced increases in plasma creatinine, dose-dependently increased renal glutathione concentration and attenuated renal lipid peroxidation as assessed by malondialdehyde levels at one week after cisplatin injection.<sup>34</sup> One hour pretreatment with a medium dose of vitamin C (100 mg/kg/day) restored the changes in urinary 8-hydroxy-2'-deoxyguanosine caused by cisplatin suggesting that vitamin C prevented oxidative DNA damage. 32 A single high vitamin C dose given six hours prior to cisplatin also prevented the decreases in the renal cortical brush border membrane enzyme (alkaline phosphatase, leucine aminopeptidase, gamma-glutamyl transferase and maltase) activities and transport of inorganic phosphate 4 days after cisplatin administration.<sup>32</sup> However, the dose of 250 mg/kg was most effective in attenuating cisplatininduced increases in plasma creatinine and BUN concentrations but lower and especially higher vitamin C doses were less effective. <sup>32</sup> In a light and electron microscopic study chronic daily treatment with vitamin C at low dose (8 mg/kg) decreased renal histological injury caused by three repeated cisplatin administrations at 21 day intervals. 31 These observations were similar to those seen in our study. Collectively we can conclude that pretreatment with vitamin C at medium-high doses of 100-250 mg/kg can attenuate cisplatin-induced nephrotoxicity.

Cisplatin elevated iron concentration in the plasma which may induce free radical reactions.<sup>50</sup> Treatment with CV247 restored the cisplatin-induced increase in plasma iron concentration. Otherwise plasma metal element content hardly differed from the control in the groups treated either with CV247 or cisplatin+CV247. The restoration of plasma iron concentration in the group treated with cisplatin+CV247 was very favourable and may explain the significant decrease of chemiluminescence intensity in the plasma.

There is always some concern whether the efficacy of a drug is impaired or not when its side effects are aimed to be reduced by administration of an adjuvant. CV247 has been

shown to have a cytotoxic effect at the G2 phase in 4 malignant human cells lines (breast, prostate, colon and lung), and CV247 produced a synergistic effect with cisplatin in 3 breast and 3 colon carcinomas by increasing the cytotoxic efficacy of cisplatin up to four-fold in cell cultures. These observations suggest that besides offering some protection from nephrotoxicity, CV247 may allow reducing the dose of cisplatin without altering its efficacy.

Cisplatin also blocks the normal accumulation of Cu and Zn in the kidney. 51 CV treatment was able to inhibit the depletion of Cu and Mn, but not of Zn.

In conclusion, the current study demonstrated that chronic CV247 administration after treatment with cisplatin offered some protection against nephrotoxicity at two weeks in rats. Since previous studies have shown that CV247 had direct toxic effects on malignant cells and also synergically increased the anticancer effect of cisplatin in cancer cell lines CV247 may enable dose reduction of cisplatin in cancer patients. A lower cisplatin dose and a partial protection against nephrotoxicity may considerably reduce the side effect of cisplatin in patients treated with CV247. However, further pharmacological studies are needed to demonstrate that CV247 also increases the anticancer effect of cisplatin in whole animal cancer models.

## Acknowledgement

The study was supported by Ivy Medical Chemicals Plc (54 Sun Street, Waltham Abbey, Essex EN9 1EJ, UK). We express our thanks to Dr Peter Karran, Head of Mammalian Cell DNA Repair Dept., Cancer Research UK, London Research Institute, Clare Hall Labs., Herts, EN6 3LD, U.K. and to Dr Anikó Bohács, Assistant Professor, Department of Pulmonology, Semmelweis University, Budapest, Hungary for their contribution.

### Figure legends

### Figure 1.

The effects of acute cisplatin and chronic CV247 administration on the body weight and daily water consumption in rats (n=10/group). Left panel: body weight. The asterisk (\*) shows that from day 3 body weight was significantly decreased in the groups treated with cisplatin and cisplatin + CV247 in comparison to the groups treated with vehicle and CV247. Right panel: water consumption. The asterisk (\*) shows that from day 5 water consumption was significantly increased in the groups treated with cisplatin and cisplatin + CV247 in comparison to the group treated with vehicle. The open circles show that on days 8, 9 and 11 water consumption was significantly increased in the group treated with CV247 in comparison to the group treated with vehicle. The statistical analysis was performed by two-way repeated measures ANOVA followed by Bonferroni post hoc test.

### Figure 2.

The effects of acute cisplatin and chronic CV247 administration on plasma creatinine and blood urea nitrogen concentration on day 12 in rats (n=10/group). CDDP increased both plasma creatinine and blood urea nitrogen (BUN) concentrations on day 12 of the experiment. CV247 did not alter plasma creatinine and BUN concentrations compared to those of the groups treated either with vehicle or with CDDP. Group effects of CDDP and CV247 and their interaction were obtained from two-way ANOVA.

### Figure 3.

The effects of acute cisplatin and chronic CV247 administration on the free radical- and reactive oxygen species (ROS)-scavenging ability of the serum of rats measured by

chemiluminescence (RLU%) on day 14 after cisplatin administration. Cisplatin increased while CV247 decreased plasma chemiluminescence. Group effects of CDDP and CV247 and their interaction were obtained from two-way ANOVA.

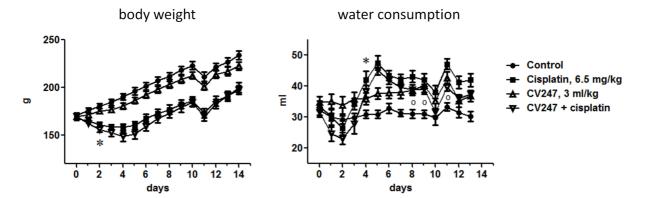
### Figure 4.

The effects of acute cisplatin and chronic CV247 administration on kidney histology (haematoxylin–eosin staining). The structure of the kidney was normal in the C (A) and CV groups (C). Severe degree of tubulointerstitial abnormality was present in rats treated with CDDP (B), and similar but significantly less severe (CDDP: 3.67±0.50 vs. CDDP+CV: 2.67±0.71; p<0.01) alterations were detected in the group treated with CDDP+CV (D). See higher magnification of sections from the group CDDP (E) and CDDP+CV (F). Blinded scores of histological abnormalities were statistically compared by two-way ANOVA.

### Figure 5.

The effects of acute cisplatin and chronic CV247 administration on COX-2 immunohistochemistry in the renal cortex. Mild activity in the interstitium and tubular epithelium was present in rats treated with vehicle (A) and CV (C). CDDP increased COX-2 activity in the damaged areas of the kidney (B), which effect was almost significantly decreased (3.00±0.71 vs. 2.44±0.53, p=0.097, Mann Whitney test) by CV (D). See higher magnification of sections from the group CDDP (E) and CDDP+CV (F). Blinded scores of COX-2 immunoreactivity were statistically compared by two-way ANOVA.

Figure 1



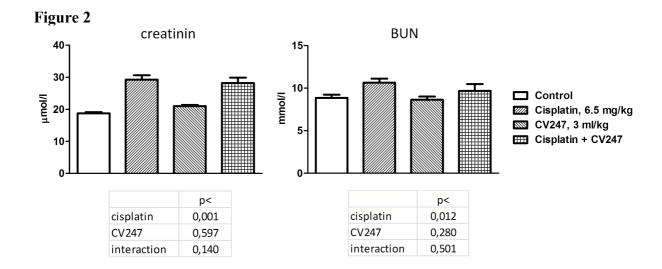
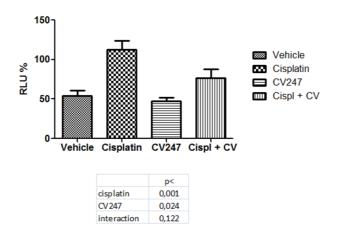
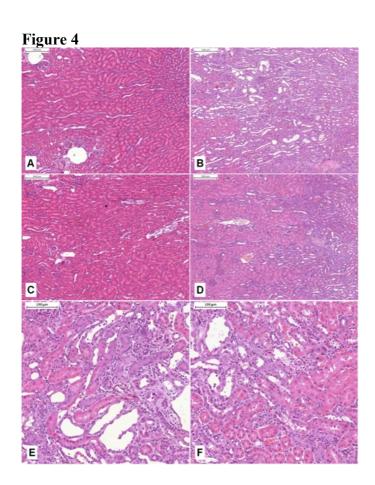
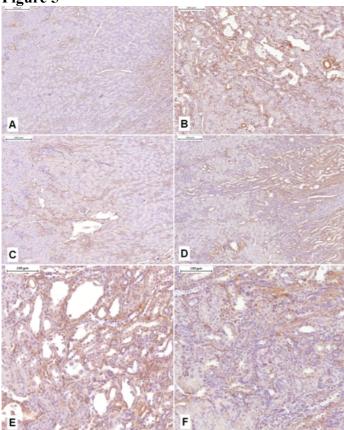


Figure 3









**Table 1.** Element concentrations of rat kidneys ( $\mu g/g$ ) measured by ICP-OES

	Element concentration					ANOVA		
	C	CDDP	CV	CDDP+CV	CDDP	CV	CDDP*CV	
	μg/g	μg/g	μg/g	μg/g	p<	p<	p<	
Cobalt (Co)	0.25±0.03	0.21±0.05	0.23±0.03	0.21±0.05	0.053	0.384	0.382	
Copper (Cu)	5.09±0.50	2.85±0.61	5.07±0.80	3.86±0.54	0.001	0.024	0.016	
Iron (Fe)	42.5±4.1	33.4±3.4	38.2±2.8	38.7±4.4	0.001	0.743	0.001	
Manganese (Mn)	0.81±0.12	0.51±0.10	0.79±0.08	$0.60\pm0.07$	0.001	0.01	0.032	
Molybdenum (Mo)	0.23±0.02	0.19±0.03	0.26±0.02	0.23±0.04	0.001	0.001	0.662	
Platinum (Pt)	BLQ	3.05±0.58	BLQ	2.07±0.25	1	0.001	-	
Selenium (Se)	0.19±0.16	0.13±0.06	0.21±0.09	0.29±0.15	0.895	0.202	0.331	
Zinc (Zn)	16.1±1.38	13.2±1.51	15.1±0.82	15.0±0.70	0.001	0.340	0.001	

BLQ: below limit of quantitation. Statistical analysis was performed

Table 2. Element content in rat plasma ( $\mu g/g$ ) measured by ICP-OES

	Element concentration					ANOVA		
	C	CDDP	CV	CDDP+CV	CDDP	CV	CDDP*CV	
	μg/g	μg/g	μg/g	μg/g	p<	p<	p<	
Copper	0.824±0.101	0.957±0.116	0.820±0.069	0.961±0.201	0.002	0.993	0.916	
Iron	3.56±0.97	5.89±1.87	3.81±1.63	3.46±0.68	0.046	0.029	0.008	
Manganese	0.026±0.012	0.041±0.025	0.024±0.012	0.029±0.016	0.082	0.208	0.378	
Molybdenu m	0.056±0.020	0.067±0.019	0.041±0.022	0.045±0.024	0.263	0.017	0.644	
Zinc	1.37±0.27	1.48±0.18	1.33±0.12	1.27±0.17	0.738	0.076	0.233	

Plasma cobalt and selenium concentrations were below limit of quantitation in most cases.

#### References

- 1. Chua DT, Ma J, Sham JS, et al. Long-term survival after cisplatin-based induction chemotherapy and radiotherapy for nasopharyngeal carcinoma: a pooled data analysis of two phase III trials. *Journal of clinical oncology: official journal of the American Society of Clinical Oncology.* 2005; 23: 1118-24.
- 2. Pignon JP, Syz N, Posner M, et al. Adjusting for patient selection suggests the addition of docetaxel to 5-fluorouracil-cisplatin induction therapy may offer survival benefit in squamous cell cancer of the head and neck. *Anti-cancer drugs*. 2004; 15: 331-40.
- 3. Rossi A, Di Maio M, Chiodini P, et al. Carboplatin- or cisplatin-based chemotherapy in first-line treatment of small-cell lung cancer: the COCIS meta-analysis of individual patient data. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2012; 30: 1692-8.
- 4. Setton J, Wolden S, Caria N and Lee N. Definitive treatment of metastatic nasopharyngeal carcinoma: Report of 5 cases with review of literature. *Head & neck.* 2012; 34: 753-7.
- 5. Rosa DD, Medeiros LR, Edelweiss MI, Pohlmann PR and Stein AT. Adjuvant platinum-based chemotherapy for early stage cervical cancer. *Cochrane database of systematic reviews (Online)*. 2012; 6: CD005342.
- 6. Boni C, Tiseo M, Boni L, et al. Triplets versus doublets, with or without cisplatin, in the first-line treatment of stage IIIB-IV non-small cell lung cancer (NSCLC) patients: a multicenter randomised factorial trial (FAST). *British journal of cancer*. 2012; 106: 658-65.
- 7. Rajeswaran A, Trojan A, Burnand B and Giannelli M. Efficacy and side effects of cisplatin- and carboplatin-based doublet chemotherapeutic regimens versus non-platinum-based doublet chemotherapeutic regimens as first line treatment of metastatic non-small cell lung carcinoma: a systematic review of randomized controlled trials. *Lung cancer (Amsterdam, Netherlands)*. 2008; 59: 1-11.
- 8. Noda K, Nishiwaki Y, Kawahara M, et al. Irinotecan plus cisplatin compared with etoposide plus cisplatin for extensive small-cell lung cancer. *The New England journal of medicine*. 2002; 346: 85-91.
- 9. Hartmann JT and Lipp HP. Toxicity of platinum compounds. *Expert opinion on pharmacotherapy*. 2003; 4: 889-901.
- 10. Yao X, Panichpisal K, Kurtzman N and Nugent K. Cisplatin nephrotoxicity: a review. *The American journal of the medical sciences*. 2007; 334: 115-24.
- 11. Miller RP, Tadagavadi RK, Ramesh G and Reeves WB. Mechanisms of Cisplatin nephrotoxicity. *Toxins*. 2010; 2: 2490-518.
- 12. Launay-Vacher V, Rey JB, Isnard-Bagnis C, Deray G and Daouphars M. Prevention of cisplatin nephrotoxicity: state of the art and recommendations from the European Society of Clinical Pharmacy Special Interest Group on Cancer Care. *Cancer chemotherapy and pharmacology*. 2008; 61: 903-9.
- 13. Pabla N and Dong Z. Cisplatin nephrotoxicity: mechanisms and renoprotective strategies. *Kidney international*. 2008; 73: 994-1007.
- 14. Lameire N, Kruse V and Rottey S. Nephrotoxicity of anticancer drugs--an underestimated problem? *Acta clinica Belgica*. 2011; 66: 337-45.
- 15. Berns JS and Ford PA. Renal toxicities of antineoplastic drugs and bone marrow transplantation. *Seminars in nephrology*. 1997; 17: 54-66.
- 16. Perazella MA and Moeckel GW. Nephrotoxicity from chemotherapeutic agents: clinical manifestations, pathobiology, and prevention/therapy. *Seminars in nephrology*. 2010; 30: 570-81.

- 17. de Jongh FE, van Veen RN, Veltman SJ, et al. Weekly high-dose cisplatin is a feasible treatment option: analysis on prognostic factors for toxicity in 400 patients. *British journal of cancer*. 2003; 88: 1199-206.
- 18. Mathe C, Bohacs A, Duffek L, et al. Cisplatin nephrotoxicity aggravated by cardiovascular disease and diabetes in lung cancer patients. *The European respiratory journal* : official journal of the European Society for Clinical Respiratory Physiology. 2011; 37: 888-94.
- 19. Safirstein R, Miller P and Guttenplan JB. Uptake and metabolism of cisplatin by rat kidney. *Kidney international*. 1984; 25: 753-8.
- 20. Pabla N, Murphy RF, Liu K and Dong Z. The copper transporter Ctr1 contributes to cisplatin uptake by renal tubular cells during cisplatin nephrotoxicity. *American journal of physiology Renal physiology*. 2009; 296: F505-11.
- 21. Yonezawa A and Inui K. Organic cation transporter OCT/SLC22A and H(+)/organic cation antiporter MATE/SLC47A are key molecules for nephrotoxicity of platinum agents. *Biochemical pharmacology*. 2011; 81: 563-8.
- 22. Katsuda H, Yamashita M, Katsura H, et al. Protecting cisplatin-induced nephrotoxicity with cimetidine does not affect antitumor activity. *Biological & pharmaceutical bulletin*. 2010; 33: 1867-71.
- 23. Filipski KK, Mathijssen RH, Mikkelsen TS, Schinkel AH and Sparreboom A. Contribution of organic cation transporter 2 (OCT2) to cisplatin-induced nephrotoxicity. *Clinical pharmacology and therapeutics*. 2009; 86: 396-402.
- 24. Chirino YI and Pedraza-Chaverri J. Role of oxidative and nitrosative stress in cisplatin-induced nephrotoxicity. *Experimental and toxicologic pathology : official journal of the Gesellschaft fur Toxikologische Pathologie.* 2009; 61: 223-42.
- 25. El-Beshbishy HA, Bahashwan SA, Aly HA and Fakher HA. Abrogation of cisplatin-induced nephrotoxicity in mice by alpha lipoic acid through ameliorating oxidative stress and enhancing gene expression of antioxidant enzymes. *European journal of pharmacology*. 2011; 668: 278-84.
- 26. Shahbazi F, Dashti-Khavidaki S, Khalili H and Lessan-Pezeshki M. Potential renoprotective effects of silymarin against nephrotoxic drugs: a review of literature. *Journal of pharmacy & pharmaceutical sciences : a publication of the Canadian Society for Pharmaceutical Sciences, Societe canadienne des sciences pharmaceutiques*. 2012; 15: 112-23.
- 27. Hensley ML, Hagerty KL, Kewalramani T, et al. American Society of Clinical Oncology 2008 clinical practice guideline update: use of chemotherapy and radiation therapy protectants. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2009; 27: 127-45.
- 28. Uzunoglu S, Karagol H, Ozpuyan F, et al. Protective effect of L-carnitine versus amifostine against cisplatin-induced nephrotoxicity in rats. *Medical oncology (Northwood, London, England)*. 2011; 28 Suppl 1: S690-6.
- 29. Weichert-Jacobsen KJ, Bannowski A, Kuppers F, Loch T and Stockle M. Direct amifostine effect on renal tubule cells in rats. *Cancer research*. 1999; 59: 3451-3.
- 30. Foster-Nora JA and Siden R. Amifostine for protection from antineoplastic drug toxicity. *American journal of health-system pharmacy : AJHP : official journal of the American Society of Health-System Pharmacists.* 1997; 54: 787-800.
- 31. Tarladacalisir YT, Kanter M and Uygun M. Protective effects of vitamin C on cisplatin-induced renal damage: a light and electron microscopic study. *Renal failure*. 2008; 30: 1-8.

- 32. Fatima S, Arivarasu NA and Mahmood R. Vitamin C attenuates cisplatin-induced alterations in renal brush border membrane enzymes and phosphate transport. *Human & experimental toxicology*. 2007; 26: 419-26.
- 33. De Martinis BS and Bianchi MD. Effect of vitamin C supplementation against cisplatin-induced toxicity and oxidative DNA damage in rats. *Pharmacological research: the official journal of the Italian Pharmacological Society.* 2001; 44: 317-20.
- 34. Antunes LM, Darin JD and Bianchi MD. Protective effects of vitamin c against cisplatin-induced nephrotoxicity and lipid peroxidation in adult rats: a dose-dependent study. *Pharmacological research: the official journal of the Italian Pharmacological Society.* 2000; 41: 405-11.
- 35. Naziroglu M, Karaoglu A and Aksoy AO. Selenium and high dose vitamin E administration protects cisplatin-induced oxidative damage to renal, liver and lens tissues in rats. *Toxicology*. 2004; 195: 221-30.
- 36. Ulubas B, Cimen MY, Apa DD, Saritas E, Muslu N and Cimen OB. The protective effects of acetylsalicylic acid on free radical production in cisplatin induced nephrotoxicity: an experimental rat model. *Drug and chemical toxicology*. 2003; 26: 259-70.
- 37. Ramesh G and Reeves WB. Salicylate reduces cisplatin nephrotoxicity by inhibition of tumor necrosis factor-alpha. *Kidney international*. 2004; 65: 490-9.
- 38. Li G, Sha SH, Zotova E, Arezzo J, Van de Water T and Schacht J. Salicylate protects hearing and kidney function from cisplatin toxicity without compromising its oncolytic action. *Laboratory investigation; a journal of technical methods and pathology*. 2002; 82: 585-96
- 39. Toloudi M, Spachidou M, Chatziioannou M, Apostolou P, Oakes R and Papasotiriou I. The Impact of CV247 Component on Human Cancer Cell Lines. *J Clin Studies* 2011; 3: 62-9.
- 40. Toloudi M, Apostolou P, Chatziioannou M, Oakes R and Papasotiriou I. Effectiveness of CDDP and CV247 combination, on colon and breast carcinomas. *J Clin Studies*. 2012; 4: 36-42.
- 41. Ognjanovic BI, Djordjevic NZ, Matic MM, et al. Lipid peroxidative damage on Cisplatin exposure and alterations in antioxidant defense system in rat kidneys: a possible protective effect of selenium. *International journal of molecular sciences*. 2012; 13: 1790-803.
- 42. Blázovics A, Kovács A, Lugasi A, Hagymási K, Biró L and Fehér J. Antioxidant defense in erythrocytes and plasma of patients with active and quiescent Crohn disease and ulcerative colitis: a chemiluminescent study. *Clinical chemistry*. 1999; 45: 895-6.
- 43. Szentmihályi K, Blázovics A, Kocsis I, Fehér E, Lakatos B and Vinkler P. The effect of fat rich diet and alcohol on ion concentration in bile fluid in rats. *Acta Alimentaria*. 2000; 29: 359-66.
- 44. May Z, Taba G, Blázovics A, et al. The application of stripping technique for the determination of Selene content in various samples with a voltammetric method. In: Majdik K, (ed.). *XIth International Chemical Conference*. Incitato Press ed.: Hungarian Technical Scientific Society of Transylvania, 2005, p. 354-6.
- 45. Verrijk R, Smolders IJ, Bosnie N and Begg AC. Reduction of systemic exposure and toxicity of cisplatin by encapsulation in poly-lactide-co-glycolide. *Cancer research*. 1992; 52: 6653-6.
- 46. Hanigan MH, Gallagher BC and Taylor PT, Jr. Cisplatin nephrotoxicity: inhibition of gamma-glutamyl transpeptidase blocks the nephrotoxicity of cisplatin without reducing platinum concentrations in the kidney. *American journal of obstetrics and gynecology*. 1996; 175: 270-3; discussion 3-4.
- 47. Blázovics A. Redox homeostasis, bioactive agents and transduction therapy. *Current Signal Transduction Therapy*. 2007; 2: 226-39.

- 48. Mitchell JA, Saunders M, Barnes PJ, Newton R and Belvisi MG. Sodium salicylate inhibits cyclo-oxygenase-2 activity independently of transcription factor (nuclear factor kappaB) activation: role of arachidonic acid. *Molecular pharmacology*. 1997; 51: 907-12.
- 49. Quilley J, Santos M and Pedraza P. Renal protective effect of chronic inhibition of COX-2 with SC-58236 in streptozotocin-diabetic rats. *American journal of physiology Heart and circulatory physiology*. 2011; 300: H2316-22.
- 50. Lasheras C, Gonzalez S, Huerta JM, Braga S, Patterson AM and Fernandez S. Plasma iron is associated with lipid peroxidation in an elderly population. *Journal of trace elements in medicine and biology : organ of the Society for Minerals and Trace Elements (GMS)*. 2003; 17: 171-6.
- 51. Mason RW and Edwards IR. Studies on the copper and zinc content of the rat kidney after treatment with cisplatin. *Toxicology*. 1985; 37: 267-74.