RESEARCH ARTICLE



WILEY

Selenate—An internal chemical shift standard for aqueous ⁷⁷Se NMR spectroscopy

Tamás Pálla	Ι	Laura Herbath	Ι	Károly Mazák	Ι	Arash Mirzahosseini 🗅	
Béla Noszál							

Department of Pharmaceutical Chemistry, Semmelweis University, Budapest, Hungary

Correspondence

Prof. Béla Noszál, Department of Pharmaceutical Chemistry, Semmelweis University, Hőgyes Endre Street 9, H-1092 Budapest, Hungary. Email: noszal.bela@pharma.semmelweisuniv.hu

Funding information

Ministry for Innovation and Technology of Hungary, Grant/Award Numbers: FIKP 2020, ÚNKP-20-4-I-SE-2, ÚNKP-20-4-II-SE-3; Ministry for Innovation and Technology

Abstract

The ⁷⁷Se NMR spectra of selenate were studied under various circumstances, such as concentration, pH, temperature, ionic strength, and D₂O:H₂O ratio, in order to examine its potential as a water-soluble internal chemical shift standard. The performance of selenate as a chemical shift reference and that of other attempted ones from the literature (dimethyl selenide, tetramethylsilane/TMS, and 3-(trimethylsilyl)propane-1-sulfonate/DSS) was also explored. The uncertainty in the resulting chemical shift relative to the effective spectral width is comparable to that of DSS. Compared to the currently prevalent water-soluble external chemical shift reference, selenic acid solution, the properties of internal selenate are much more favorable in terms of ease of use. We have also demonstrated that selenate can be used in reducing media, which is inevitable for the analysis of selenol compounds. Thus, it can be stated that sodium selenate is a robust internal chemical shift reference in aqueous media for ⁷⁷Se NMR measurements; the chemical shift of this reference in a solution containing 5 V/V% D_2O at 25°C and 0.15 mol·dm⁻³ ionic strength is 1048.65 ppm relative to 60 V/V% dimethyl selenide in CDCl₃ and 1046.40 ppm relative to the ¹H signal of 0.03 V/V% TMS in CDCl₃. In summary, a water-soluble, selenium-containing internal chemical shift reference compound was introduced for ⁷⁷Se NMR measurements for the first time in the literature, and with the aforementioned results all previous ⁷⁷Se measurements can be converted to a unified scale defined by the International Union of Pure and Applied Chemistry.

KEYWORDS

chemical shift standard, ppm reference, selenium NMR

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. Magnetic Resonance in Chemistry published by John Wiley & Sons Ltd.

1 | INTRODUCTION

The nucleus of ⁷⁷Se isotope has valuable, yet underutilized properties for nuclear magnetic resonance spectroscopy (spin: 1/2; natural abundance: 7.63%; relative receptivity: $5.37 \cdot 10^{-4}$ with respect to ¹H and 3.15 with respect to ¹³C; chemical shift range: \sim 2,800 ppm). The availability of ⁷⁷Se NMR spectroscopy was demonstrated in the early 1950s,^[1,2] then it quickly evolved as a selective method to investigate selenium-containing compounds.^[3-6] The biological significance of seleniumcontaining proteins (selenoproteins) was described early on in the maintenance of physiological redox homeostasis and it has also been proven that the selenocysteine residues are essential for the catalytic activity of such redox enzymes.^[7-10] Moreover, our research group has recently found close correlation among the standard redox potential, the acid-base properties and the ⁷⁷Se chemical shift of selenolate-diselenide redox systems.^[11,12] These findings will no doubt lead to a more extensive use of ⁷⁷Se NMR as a selective analytical tool in the characterization of selenoproteins and organoselenium compounds. Despite the long-standing use of this technique, a reliable and robust chemical shift standard is still an unmet need, as seen in a systematic error between separately reported ⁷⁷Se chemical shifts in the literature.^[12]

In the ⁷⁷Se NMR literature, chemical shift data are referenced to a relatively large number of compounds(seleninyl chloride,^[13] dimethyl selenide,^[14,15] diphenyl diselenide,^[20-24] selenophene,^[16–19] 4.4'dimethyldiphenyl diselenide,^[25] selenium oxide,^[26] sodium selenite,^[27] selenous acid).^[13,28-31] In addition, indication of the measurement parameters (concentration, ionic strength, temperature) is rather the exception than the case. This may cause differences and misunderstanding between various results. It is noteworthy that selenic acid has already been used as a chemical shift reference^[13]; however, the measurement conditions were not specified. As we have shown certain measurement parameters can significantly affect the frequency of the selenate signal, the precise knowledge of measurement circumstances can therefore have a crucial effect on the research findings.

There are two compounds that are commonly used as chemical shift reference for ⁷⁷Se NMR, namely dimethyl selenide^[14] and selenous acid.^[32] The solvent-, concentration- and temperature-dependence of ⁷⁷Se chemical shift of dimethyl selenide has been elaborated. Based on literature recommendation, in principle, this reference can be used as an internal standard in nonpolar solvents; however, it is virtually always used as an external reference compound. The disadvantages of this compound are that dimethyl selenide is not water soluble; therefore, the external standard method will cause some inconveniency in aqueous measurements. Furthermore, dimethyl selenide is an extremely volatile and toxic substance.

Selenous acid is only recommended as an external reference compound and the temperature-, concentration-, solvent- and pH-dependence of the ⁷⁷Se chemical shift has been elaborated. However, in every work using selenous acid as chemical shift standard the final chemical shift data are referred to 60 V/V%dimethyl selenide in CDCl₃. Since selenite undergoes two protonation processes ($\log K_1 = 8.20 \pm 0.01$ and $\log K_2 = 2.46 \pm 0.06$, these values are in good agreement of the previously described parameters^[33]), it is certainly unfit to be used as a reliable internal chemical shift reference in aqueous media where pH changes usually take place. Given the obvious requirements of an internal standard, a reliable and robust ⁷⁷Se NMR chemical shift reference compound which can be used in aqueous media is a definite precondition for future selenoprotein analysis. Other potential reference compounds from the available organoselenium compounds were thoroughly investigated and excluded due to poor water solubility, high toxicity or having a protonating group in the pH range of interest. Notably, N,N-dimethylselenourea was excluded because of having an acid-base function $(\log K = 4.98 \pm 0.01)$, just like dimethyl selenoxide $(\log K = 2.04 \pm 0.03)$, in agreement with previous findings^[34]) which is also unstable and hygroscopic in pure form.

In this work we introduce selenate and demonstrate its properties as an internal ⁷⁷Se chemical shift reference for aqueous media. The pH-, temperature-, concentration-, ionic strength-, and solvent-dependence of the ⁷⁷Se frequency are analyzed in terms of a multilinear regression model. Its chemical stability and reactivity in various media were also determined.

2 | MATERIALS AND METHODS

2.1 | Materials

Selenic acid (40 m/m%), anhydrous sodium selenate, selenous acid, N,N-dimethylselenourea, dimethyl selenoxide, sodium chloride and seleno-L-cystine were purchased from Sigma (Merck), sodium 3-(trimethylsilyl)-1-propanesulfonate (DSS) and dimethyl selenide were obtained from Tokyo Chemical Industry, deuterium oxide (D₂O) and chloroform-d containing 0.03 V/V% tetramethylsilane (TMS) were purchased from Merck. All the above-mentioned substances were used without further purification. Deionized water was prepared with a Milli-Q Direct 8 Millipore system.

2.2 | NMR spectroscopy measurements

NMR spectra were recorded on a Varian Unity Inova DDR spectrometer (599.9 MHz for ¹H) with a 5 mm ¹H{¹³C/³¹P-¹⁵N} pulse field gradient triple resonance probehead at 298.15 \pm 0.1 K. The solvent was H₂O: D₂O 95:5 (*V/V*) unless otherwise stated, ionic strength was adjusted to either 0.15 or 1.0 mol·dm⁻³ unless otherwise stated. The concentration of sodium selenate varied between 0.0025 and 0.5 mol·dm⁻³. For the stability tests of selenate two samples with different ionic strengths and pH were prepared and stored at room temperature. The ⁷⁷Se NMR spectra of these samples were recorded at different time points between 1 minute and 130 days. The experimental design to study the above mentioned parameters is presented in Table 1.

The pH values were determined in situ by internal indicator molecules (at ca. 1 mmol·dm⁻³) optimized for ¹H NMR.^[35,36] The sample volume was 600 μ l and every sample contained ca. 1 mmol·dm⁻³ DSS. Where necessary, the H₂O ¹H signal was suppressed with a presaturation sequence; the average acquisition parameters for ¹H measurements are: number of transients = 16, number of points = 65,536, acquisition time = 3.33 s, relaxation delay = 1.5 s. The ⁷⁷Se NMR parameters were as follows: pulse and acquire sequence

using 7.25 μ s observe pulse, at most the number of transients was between 128 and 65,536 based on the concentration of the selenium-containing compound, number of points = 262,144, maximal spectral width applied = 131578.9 Hz, relaxation delay = 1 s. The ⁷⁷Se measurements were acquired with a standard s2pul pulse sequence optimized for the ⁷⁷Se nucleus at pulse width = 14.50 μ s (at 60 dB) with ¹H decoupling during the pulse and FID acquisition. The presence or absence of the ¹H decoupling had no effect on the signal frequency or width.

Based on IUPAC recommendations^[37,38] the resulting ⁷⁷Se NMR spectra were referenced to the ¹H NMR signal of TMS (measured in a separate sample at 0.03 V/V% in CDCl₃) using the *substitution method*. However, the chemical shift referencing was also carried out relative to the ¹H NMR signal of DSS methyl groups present in the same sample using the *substitution method*, in order to determine whether this widely accepted ¹H reference can be used instead of TMS.

Multiple linear regression was carried out by MedCalc 12.5.00 software (MedCalc Software Ltd., Ostend, Belgium).

2.3 | pH-potentiometric titrations

A 716 DMS Titrino automatic titrator (Metrohm) with a Metrohm 6.0234.110 combined pH glass electrode was used for the potentiometric titrations, under automatic PC (personal computer) control. The pH-potentiometric

TABLE 1 The investigated parameters and the exact measurement parameters (each line corresponding to an investigated parameter) applied to characterize selenic acid ⁷⁷Se chemical shifts

		Other measurement parameters				
Investigated parameter	Investigated range	pН	Т (К)	Selenate <i>c</i> (mol∙dm ⁻³)	I (mol∙dm ⁻³)	<i>V/V</i> % D ₂ O
pH	1.36-11.68	NA	298	0.05	0.15	5
	0.66-13.09	NA	298	0.333	1.00	5
<i>T</i> (K)	293–328 (6 measurement points)	8.78	NA	0.333	1.00	5
selenate c (mol·dm ⁻³)	0.0025–0.5 (9 measurement points)	~7	298	NA	1.5	5
$I (\mathrm{mol} \cdot \mathrm{dm}^{-3})$	0.15–1 (7 measurement points)	~7	298	0.05	NA	5
<i>V/V</i> % D ₂ O	5–100 (11 measurement points)	~7	298	0.333	1.00	NA
stability	stability observed throughout 130 days	11.50	298	0.05	0.15	5
		8.78	298	0.333	1.00	5

Note. pH \sim 7 refers to the native pH of aqueous solutions of sodium selenate and sodium chloride.

system was calibrated using pH 1.68, 4.01, 6.87, 9.18 aqueous National Bureau of Standards (NBS) buffer solutions. Constant temperature $(25 \pm 0.1^{\circ}C)$ was provided by a thermostated double-walled glass cell. Difference titrations were carried out in the absence (blank) and presence of ligands. First 2 ml of 0.1 mol·dm⁻³ HCl was titrated with 0.1 mol·dm⁻³ KOH. Constant ionic strength of 0.15 mol·dm⁻³ was provided by the presence of KCl. Then, a ligand was added to the same volume of HCl solution and was subsequently titrated with KOH. The initial concentration of the ligands varied between 10 and 20 mmol·dm⁻³ in the titrations. Nonlinear parameter fitting with Origin 8 provided the protonation constants from the volume differences.

3 | RESULTS

3.1 | pH dependence

Protonation of selenate significantly changes the electron structure of the species and therefore has a high perturbing effect on the chemical shift modelled described by the following equation:

$$\delta_{\rm obs}(\rm pH) = \frac{\delta_{\rm L} + \sum_{i=1}^{2} \delta_{\rm H_iL} \times 10^{\log\beta_i - i \cdot \rm pH}}{1 + \sum_{i=1}^{2} 10^{\log\beta_i - i \cdot \rm pH}}, \qquad (1)$$

where $\delta_{\rm L}$ is the chemical shift of an unprotonated ligand (L), $\delta_{\rm H_iL}$ values stand for the chemical shifts of successively protonated ligands, and $\beta_{\rm i}$ is the cumulative protonation macroconstant; log henceforth refers to the

base 10 logarithm. Note that under normal aqueous conditions (0 < pH < 14) only one protonation step of selenate takes place, thus in such a monoprotic case, in Equation 1 i = {1}.

However, as selenic acid is a strong diprotic acid (even stronger than sulfuric acid), even the first protonation step of selenate ions only takes part below pH = 3.5. To obtain the sensitivity of ⁷⁷Se nuclei in selenic acid to acid-base processes, the complete ⁷⁷Se-NMR pH titration was carried out at two different ionic strengths, namely, 0.15 mol·dm⁻³ and 1 mol \cdot dm⁻³; the concentration of selenate ions was $0.05 \text{ mol} \cdot \text{dm}^{-3}$ and $0.33 \text{ mol} \cdot \text{dm}^{-3}$, respectively. The measured samples all contained DSS, therefore the ¹H methyl DSS chemical shifts were also observed relative to TMS; the resulting titration curves are shown in Figure 1. We did not fit a titration curve onto the obtained data points in order to determine the protonation constants, since the low pH plateaus of the first protonation steps are not visible under the measurement conditions. Note that the ¹H methyl DSS signal chemical shift varies below pH 3.5 only because of the changing water resonance frequency (from previous results not shown: the H₂O ¹H chemical shift changes considerably below pH 2 and slightly above pH 13). Since the substitution method calculation makes use of the solvent deuterium frequency, it is expected that the otherwise pH-insensitive DSS signal will show a change in chemical shift when referred to TMS. In order to demonstrate the pH-insensitivity of DSS, an experiment was carried out where acetone and DSS were titrated together and the stable chemical shift of acetone can be seen when referred to internal DSS (Figure 1).

FIGURE 1 Visible in black, the pHdependence of ⁷⁷Se and ¹H (methyl) TMSreferred chemical shifts of selenate and DSS, respectively. Solid plot points were recorded at 1 mol·dm⁻³, while empty plot points were recorded at 0.15 mol·dm^{-3} ionic strength. Visible in blue, the pH-dependence of ⁷⁷Se DSSreferred chemical shifts of selenate, using the DSS within each sample. Solid plot points were recorded at 1 mol·dm⁻³, while empty plot points were recorded at 0.15 mol·dm⁻³ ionic strength. Crossed empty points show the DSS-referred ¹H chemical shifts of acetone in a pH titration experiment under the same conditions as the above



¹⁵² WILEY

3.2 | Temperature

The temperature dependence of the ⁷⁷Se chemical shift of selenate⁻ was studied from 293 to 328 K (Figure 2). In the knowledge of pH dependence, a slightly basic medium was chosen to avoid the perturbing effect of pH. The concentration of the selenate ions was $0.33 \text{ mol}\cdot\text{dm}^{-3}$ at 1.0 mol·dm⁻³ ionic strength.

3.3 | Ionic strength

The ⁷⁷Se and ¹H chemical shift values were recorded for solutions with different ionic strength at 0.15 and

1.0 mol·dm⁻³ and constant selenate concentration (0.05 mol·dm⁻³). The results are shown in Figure 3.

3.4 | Concentration

The variation of analyte concentration has a relatively small effect on the 77 Se chemical shift of selenate and likewise on the 1 H (methyl) chemical shift of DSS; the result of the measurements is plotted in Figure 4. Two stock solutions, one with 0.5 mol·dm⁻³ sodium selenate and 1 mmol·dm⁻³ DSS, the latter with 1.5 mol·dm⁻³ sodium chloride, were used to obtain samples with different selenate and DSS

FIGURE 2 The temperature dependence of ⁷⁷Se and ¹H (methyl) chemical shifts of selenic acid and DSS, respectively. The chemical shift values were referred to the ¹H signal of 0.03 V/V % TMS in CDCl₃





concentration at a constant ionic strength of $1.5 \text{ mol} \cdot \text{dm}^{-3}$.

significant effect on the chemical shift. D_2O content of the solvent mixture was changed from 5 to 100 *V*/*V*%; the results are plotted in Figure 5.

3.5 | D_2O saturation

Water and deuterium oxide are similar in their main physico-chemical properties; however, due to their different relative permittivity values, they can exert a perturbing effect on the chemical shifts. Thus, H_2O/D_2O mixtures with different component ratios may have a

3.6 | Stability

Selenate-containing solutions at two different ionic strengths in slightly basic media were stored in NMR tubes at room temperature for more than 4 months. The effect of the time elapsed after preparing the solutions on



FIGURE 4 The concentration dependence of ⁷⁷Se and ¹H (methyl) chemical shifts of selenate and DSS, respectively. The chemical shift values were referred to the ¹H signal of 0.03 V/V% TMS in CDCl₃

FIGURE 5 The effect of the D_2O content of the solvent on the ⁷⁷Se chemical shifts of selenate. The chemical shift values were referred to the ¹H signal of 0.03 V/V% TMS in CDCl₃

the ⁷⁷Se chemical shift was one order of magnitude lower than that of any other measurement parameter.

3.7 | Redox reactivity

Since selenium-containing proteins and amino acids are sensitive to oxidation by air, it is necessary to ensure a reductive medium during their analysis. This is usually achieved by adding a reducing agent like dithiothreitol (DTT) to the mixture. Selenium is at the most oxidized (+6) state in selenate ion. In a reductive medium with DTT an undesirable side-reaction may occur, disrupting NMR measurements; namely, selenate may turn into a reduced state, for example, selenite. To demonstrate the resistance of selenate to reduction by DTT, a solution containing a diselenide compound (selenocystine), DTT as reducing agent, and selenate were investigated (Figure 6). Based on the spectra no selenite was detectable, while DTT reduced selenocystine to its reduced form, selenocysteine. Thus selenate is a stable, suitable chemical shift reference for selenopeptide measurements, even in the presence of DTT.

4 | DISCUSSION

All of the recorded ⁷⁷Se and ¹H (methyl) chemical shift values of selenate and DSS are collated in Table 2. In order to compare the utility of selenate as an NMR chemical shift reference its relative uncertainty in chemical shift given as a percentage of the ⁷⁷Se chemical shift range (2,800 ppm) was compared to that of DSS and the ¹H chemical shift range (46 ppm).



FIGURE 6 ⁷⁷Se NMR spectrum of a solution containing selenocystine, dithiothreitol and sodium selenate. The reduction of the diselenide bridge was successful as the NMR signal of the diselenide is missing at ca. 250 ppm.^[12] The lack of a selenite ⁷⁷Se NMR signal at 1200 ppm. Milne^[32] indicates the redox stability of selenate under such media

Parameter	Range investigated	$\Delta \delta^{77_{Se}}_{SeO_4^{2^-}}$	Relative error	$\Delta \delta^{1_{ m H}}_{ m DSS}$	Relative error	
T (K)	293-328	1.49	0.053%	0.29	0.63%	
$c (\mathrm{mol}\cdot\mathrm{dm}^{-3})$	0.0025-0.5	0.46	0.016%	0.10	0.22%	
$I (\mathrm{mol}\cdot\mathrm{dm}^{-3})$	0.15–1	0.59	0.021%	0.08	0.17%	
pН	1.36–11.68	2.78	0.099%	0.03	0.07%	
	0.66-13.09	4.97	0.178%	0.14	0.30%	
<i>V/V</i> % D ₂ O	5-100	0.35	0.013%	0.02	0.04%	
t (days)	127	0.05	0.002%	0.02	0.04%	
	133	0.08	0.003%	0.01	0.02%	
Reliable pH intervals						
pH range	Ionic strength	$\Delta \delta^{77_{Se}}_{SeO_4^{2-}}$	Relative error	$\Delta \delta_{DSS}^{1_{\rm H}}$	Relative error	
3.40-11.68	$I = 0.15 \text{ mol} \cdot \text{dm}^{-3}$	0.03	0.001%	0.04	0.09%	
3.55-13.09	$I = 1.0 \text{ mol} \cdot \text{dm}^{-3}$	0.11	0.004%	0.11	0.24%	

TABLE 2 The chemical shift change of ⁷⁷Se and ¹H (methyl) chemical shift of selenate and DSS, respectively, upon varying certain measurement parameters in the given ranges

Note. The relative error is calculated by expressing the chemical shift change as a percentage relative to the typical NMR chemical shift range of the nucleus in question. The data above pH 3.5 are highlighted as reliable pH intervals, as this is the interval where pH has no significant effect on the chemical shift of the reference.

A multiple linear regression model was fitted to the data above pH 3.5 as an acceptable approximation for a deterministic model to investigate the significant measurement parameters (Table 3). Using a significance level of 0.05 it was revealed that the following parameters had no significant contribution to the linear model above pH 3.5: pH, selenate concentration and time elapsed after sample preparation.

Neglecting the parameters with insignificant perturbing effects, the following equation arises to determine selenate ⁷⁷Se chemical shifts in any custom measuring environment falling into our range of investigation:

$$\delta_{\text{TMS}} = 1034.8 + 0.0390 \cdot T(\text{K}) + 0.434 \cdot I(\text{mol} \cdot \text{dm}^{-3}) + 0.0019 \cdot V/V\%\text{D}_2\text{O}$$
(2)

The value of the adjusted coefficient of determination $(R^2 = 0.8935)$ and the fact that every measured ⁷⁷Se chemical shift value was reproducible with a maximum of 0.22 ppm error also verify the results. The random errors of determination showed a normal distribution.

In order to achieve consistent chemical shift data with previous literature as well, the chemical shift of selenate was determined with respect to the more commonly used ⁷⁷Se chemical shift standards, such as TMS, DSS and dimethyl selenide. The results are shown in Table 4.

The ⁷⁷Se chemical shift of selenate ions in aqueous solution was investigated by changing six measurement parameters. Of these six parameters the pH proved to have the largest effect on the selenate signal, however the selenate protonation occurs with a logK < 2. Therefore the effect of pH on the signal is insignificant above pH = 3.5; at lower pH values the expected chemical shift can be predicted based on the titration curves. It is noteworthy that the most commonly used aqueous chemical shift reference, DSS, also undergoes protonation at such pH values. The chemical shift error caused by

TABLE 3 Results of the multiple linear regression applied to data at pH > 3.5

Constant	1,034,795		
parameter	coefficient	standard deviation	<i>p</i> value
<i>T</i> (K)	0.0390	0.0028	< 0.0001
$c (\mathrm{mol}\cdot\mathrm{dm}^{-3})$	-0.080	0.088	0.3626
$I (\mathrm{mol} \cdot \mathrm{dm}^{-3})$	0.434	0.033	< 0.0001
рН	-0.0057	0.0074	0.4493
<i>V/V</i> % D ₂ O	0.00193	0.00063	0.0033
t (days)	-0.00036	0.00049	0.4678

Note. The parameters with a *p* value lower than 0.05 were accepted as statistically significant.

TABLE 4 ⁷⁷Se NMR chemical shift of selenate ions (5 V/V% D_2O , T = 25°C, I = 0.15 mol·dm⁻³) with respect to the currently used chemical shift references for ¹H and ⁷⁷Se

⁷⁷ Se NMR chemical shift of selenate with respect to					
¹ H signal of 0.03 <i>V/V</i> % TMS in CDCl ₃	¹ H methyl signal of 1 mmol∙dm ⁻³ DSS in H ₂ O containing 5% D ₂ O	⁷⁷ Se signal of 60 <i>V/V</i> % dimethyl selenide in CDCl ₃			
1046.40 ppm	1043.96 ppm	1048.65 ppm			

protonation relative to the spectral width is comparable for the two materials.

The temperature coefficient in a linear regression model was found to be 0.042 ppm/K while keeping other parameters constant; on the other hand in a multivariate statistical analysis it proved to be 0.039 ppm/K. This value is in agreement with those of the two ⁷⁷Se chemical shifts introduced so far, i.e. dimethyl selenide $(0.025 \text{ ppm/K})^{[14]}$ and selenous acid $(0.094 \text{ ppm/K})^{[32]}$ The concentration, ionic strength and D₂O content of the solvent affect the selenate chemical shift by one order of magnitude lower, compared to the two measurement conditions mentioned above. Based on a multiple linear regression analysis, the selenate concentration is not a significant parameter, while ionic strength and solvent composition have a significant effect in the model.

Selenate ion-containing aqueous solutions at various ionic strengths and extreme basic pH values were stored at room temperature for 130 days, under which the ⁷⁷Se NMR spectrum showed no change. Compared to the currently known water-soluble reference, selenous acid, the properties of selenate are much more favorable. It has been shown experimentally that selenate salts can be used even in reducing media required for the analysis of diselenide/selenol compounds, since selenate reacts neither with thiol-containing reducing agents nor with the selenium-containing substance to be measured. Compared to the uncertainty in DSS methyl chemical shift relative to the common ¹H spectral width, the uncertainty of selenate chemical shift relative to the common ⁷⁷Se spectral width is comparable or lower in case of each six investigated measurement parameter.

5 | CONCLUSION

In this work sodium selenate was introduced as an internal chemical shift standard for ⁷⁷Se NMR measurement of aqueous solutions. The perturbing effect of pH, temperature, ionic strength, selenate concentration and D_2O content on the chemical shift was investigated. Selenate \perp Wiley-

proved to be a suitable internal chemical shift reference above pH 3.5, where the following parameters have significant effect on the chemical shift based on a multiple linear regression model: temperature, ionic strength and D₂O content. Moreover, selenate was found to be stable under the reductive conditions of dithiothreitol, the strong reducing agent, therefore selenate is suitable for investigation of diselenide compounds that require reduction prior measurement. Based on our results, sodium selenate is the most suitable internal chemical shift reference for aqueous ⁷⁷Se NMR in the literature.

ACKNOWLEDGEMENTS

This work was supported by FIKP 2020 and the ÚNKP-20-4-I-SE-2 and ÚNKP-20-4-II-SE-3 New National Excellence Program of the Ministry for Innovation and Technology of Hungary from the source of the National Research, Development and Innovation Fund, The invaluable discussions with Dr. Zoltán Szakács are much appreciated.

PEER REVIEW

The peer review history for this article is available at https://publons.com/publon/10.1002/mrc.5196.

ORCID

Arash Mirzahosseini D https://orcid.org/0000-0002-3281-8435

REFERENCES

- [1] S. Dharmatti, H. Weaver Jr., Phys. Rev. 1952, 86(2), 259.
- [2] H. Walchli, Phys. Rev. 1953, 90(2), 331.
- [3] M. Lardon, J. Am. Chem. Soc. 1970, 92(17), 5063.
- [4] G. Ribaudo, M. Bellanda, I. Menegazzo, L. P. Wolters, M. Bortoli, G. Ferrer-Sueta, G. Zagotto, L. Orian, Chemistry-A European Journal 2017, 23(10), 2405.
- [5] K. N. Sands, E. Mendoza Rengifo, G. N. George, I. J. Pickering, B. S. Gelfand, T. G. Back, Angewandte Chemie International Edition 2020, 59(11), 4283.
- [6] M. S. Silva, D. Alves, D. Hartwig, R. G. Jacob, G. Perin, E. J. Lenardão, Asian J. Org. Chem. 2021, 10(1), 91.
- [7] L. Flohe, W. Günzler, H. Schock, FEBS Lett. 1973, 32(1), 132.
- [8] J. Köhrle, Cellular and Molecular Life Sciences CMLS 2000, 57(13-14), 1853.
- [9] H. Tapiero, D. M. Townsend, K. D. Tew, Biomed. Pharmacother. 2003, 57(3), 134.
- [10] L. Zhong, E. S. Arnér, A. Holmgren, Proceedings of the National Academy of Sciences 2000, 97(11), 5854.
- [11] T. Pálla, A. Mirzahosseini, B. Noszál, Antioxidants 2020, 9(6), 465.
- [12] T. Pálla, A. Mirzahosseini, B. Noszál, Chem. Phys. Lett. 2020, 741, 137076.
- [13] T. Birchall, R. Gillespie, S. Vekris, Can. J. Chem. 1965, 43(6), 1672.
- [14] N. Luthra, R. Dunlap, J. Odom, Journal of Magnetic Resonance (1969) 1983, 52(2), 318.

- [15] W. McFarlane, R. Wood, Journal of the Chemical Society, Dalton Transactions 1972, 13, 1397.
- [16] A. Fredga, S. Gronowitz, A. B. Hörnfeldt, Chemischer Informationsdienst 1975, 6(43). https://doi.org/10.1002/chin. 197543358
- [17] S. Gronowitz, I. Johnson, A. B. Hörnfeldt, Chemischer Informationsdienst 1975, 6(22). https://doi.org/10.1002/chin. 197522079
- [18] S. Gronowitz, I. Johnson, A. B. Hornfeldt, Chemica Scripta 1975, 8(1), 8.
- [19] S. Gronowitz, A. Konar, A. B. Hörnfeldt, Organic Magnetic Resonance 1977, 9(4), 213.
- [20] S. Malik, H. Duddeck, J. Omelanczuk, M. I. Choudhary, Chirality: The Pharmacological, Biological, and Chemical Consequences of Molecular Asymmetry 2002, 14(5), 407.
- [21] P. Mecik, B. Pigulski, S. Szafert, Org. Lett. 2021, 23(3), 1066.
- [22] P. Nobre, T. Peglow, R. Bartz, A. Barcellos, R. Jacob, M. Silva, T. Barcellos, G. Perin, J. Braz. Chem. Soc. 2021. https://doi.org/ 10.21577/0103-5053.20210051
- [23] A. K. O. Silva, F. C. Pinto, K. M. Canuto, R. Braz-Filho, R. A. C. Silva, F. A. Santos, N. K. V. Monteiro, E. R. Silveira, O. D. L. Pessoa, J. Braz. Chem. Soc. 2021, 32(7), 1424.
- [24] A. Sørensen, B. Rasmussen, M. Pittelkow, The Journal of Organic Chemistry 2015, 80(8), 3852.
- [25] B. Kohne, W. Lohner, K. Praefcke, H. J. Jakobsen, B. Villadsen, J. Organomet. Chem. 1979, 166(3), 373.
- [26] P. Pekonen, Y. Hiltunen, R. S. Laitinen, T. A. Pakkanen, Inorg. Chem. 1990, 29(15), 2770.
- [27] W. Koch, O. Lutz, A. Nolle, Zeitschrift für Physik A Atoms and Nuclei 1978, 289(1), 17.
- [28] B. Günther, O. Kanert, Phys. Rev. B 1985, 31(1), 20.
- [29] M. Lamoureux, J. Milne, Polyhedron 1990, 9(4), 589.
- [30] D. L. Rabenstein, K. S. Tan, Magn. Reson. Chem. 1988, 26(12), 1079.
- [31] K.-S. Tan, A. P. Arnold, D. L. Rabenstein, Can. J. Chem. 1988, 66(1), 54.
- [32] J. Milne, Magn. Reson. Chem. 1993, 31(7), 652.
- [33] L. Barcza, L. Sillen, Acta Chemica Scandinavica 1971, 25(4), 1250.
- [34] P. Nylén, Zeitschrift für anorganische und allgemeine Chemie 1941, 246(3), 227.
- [35] G. Orgovan, B. Noszal, J. Pharm. Biomed. Anal. 2011, 54(5), 958.
- [36] Z. Szakács, G. Hägele, R. Tyka, Anal. Chim. Acta 2004, 522(2), 247.
- [37] R. K. Harris, E. D. Becker, S. M. C. De Menezes, R. Goodfellow, P. Granger, Pure and Applied Chemistry 2001, 73(11), 1795.
- [38] R. K. Harris, E. D. Becker, S. M. C. De Menezes, P. Granger, R. E. Hoffman, K. W. Zilm, Pure and Applied Chemistry 2008, 80(1), 59.

How to cite this article: T. Pálla, L. Herbath, K. Mazák, A. Mirzahosseini, B. Noszál, Magn Reson Chem 2022, 60(1), 148. https://doi.org/10. 1002/mrc.5196

156